



Antioxidant activity of *Malpighia glabra* L., leaves extract.

Amany M. Fekry^{1*}, Walaa A El-sabbagh¹, Marwa S. Abu Bakr², Mona A. EL-Ghazaly¹, Abd El-Salam I. Mohammed³

¹ Department of Drug Radiation Research, National Center for Radiation Research and Technology (NCRRT), Egyptian Atomic Energy Authority, Cairo, Egypt

² Department of Pharmacognosy, Faculty of Pharmacy (Girls), Al-Azhar University, Cairo, Egypt

³ Department of Pharmacognosy, Faculty of Pharmacy (Boys), Al-Azhar University, Cairo, Egypt

*Correspondence e-mail, a.fekry.sallam89@gmail.com

Article history: Received 2021-01-26

Revised 2021-03-24

Accepted 2021-04-05

Abstract: *Malpighia glabra*. L (Acerola) fruits have wide global medicinal uses due to rich antioxidant contents. Little studies were carried on other parts of plant. *Malpighia glabra*. L leaves (1.5 kg) was subjected to soxhlet exhaustive extraction with methanol and concentrated under vacuum. The dried extract was defatted with petroleum-ether then fractionated with ethyl acetate & n-butanol 3 successive times respectively. The ethyl acetate fraction (9 gm) was subjected to series of silica gel & Sephadex (LH-20) column chromatography. Phytochemical investigation of the Ethyl acetate fraction of *Malpighia glabra*. L leaves in this study resulted in separation of four pure compounds (Caffeic acid, Chlorogenic acid, Quercetin & Kaempferol). The structures of the separated compounds were assigned based on NMR spectral data and mass spectrometer analysis. The methanolic extract showed high values of total phenolics (96.36 mg GAE/g), flavonoids (30.71 mg QE/g) and higher antioxidant activity towards DPPH radical with $IC_{50} = 49.8(\text{mg/ml})$.

Keywords: *Malpighia glabra* L. Leaves; DPPH; Caffeic Acid; Chlorogenic Acid; Quercetin; Kaempferol

1. INTRODUCTION

Folk medicine is known to have a great influence on human culture, so that tracing and identifying of active constituents phytochemically and their medicinal effects have taken a wide interest overtime¹.

Bioactive compounds present naturally in herbs, foods and dietary supplements that have therapeutic effects and may help in protection from chronic diseases known as "Nutraceuticals"².

Malpighiaceae family have about 40-50 species, *Malpighia glabra*. L genus belongs to this family and known by other terms like Acerola, West Indian cherry, *Malpighia punicifolia* L, *Malpighia emarginata* DC and Barbados cherry^{3,4}.

Brazil and tropical countries use *Malpighia glabra*. L fruit juice as one of the highest levels of ascorbic acid, vitamin C and abundance of carotenoids and anthocyanins⁵.

Bioactive compounds which separated from *Malpighia glabra*. L in other studies shows antioxidant, antidiabetic and skin protecting activity.⁶

2. MATERIALS AND METHODS

2.1. Plant material

Malpighia glabra.L leaves were collected from Orman garden, Giza, Egypt. during May/June 2015.

2.2. Extraction and isolation

Air dried powder of *Malpighia glabra*. L leaves (1.5 kg) was exhaustively extracted with the 99% methanol (3x 10 L) using soxhlet apparatus. The combined extract by methanol was dried and conc. at 40°C under vacuum to give 450 gm of dried extract. The concentrated methanolic extract was dissolved in (500 ml) of distilled water and defatted with petroleum-ether (40 – 60°C). The crude defatted extract (150 gm) was fractionated with ethyl acetate & n-butanol respectively 3 successive times (500 ml). The ethyl acetate (9 gm) fraction was subjected to series of silica gel (Si gel 60, Sigma aldrich) and Sephadex LH-20 (Sigma aldrich) column chromatography to afford four pure compounds.

2.3. Phytochemical screening

The methanolic extract of *Malpighia glabra*. L leaves was phytochemically studied for their

Cite this article: Fekry, A., Elsabbagh, W., Abu Bakr, M., El-Ghazaly, M., Mohamed, A. Antioxidant activity of *Malpighia glabra* L., leaves extract. Azhar International Journal of Pharmaceutical and Medical Sciences, 2021; 1(2):88-93. doi: 10.21608/aijpm.2021.59935.1042

DOI: 10.21608/aijpm.2021.59935.1042

<https://aijpm.journals.ekb.eg/>

chemical constituents according to the standard methods) ⁸⁻¹¹.

2.4. Quantification of total phenolic content

Content of phenolics in Malpighia extract were determined Spectrophotometrically ¹². (The phenolics concentration was measured from the calibration curve as (mg/ml); then the phenolic content in extracts was expressed as (mg of GAE/g of extract) ¹².

2.5. Quantification of total flavonoid content

Contents of total flavonoids were assayed by the aluminum chloride colorimetric method at 510 nm ¹³.

(The total flavonoidal content was expressed in terms of mg quercetin equivalents (QE)/g dry basis) ¹³.

2.6. Antioxidant assay

Scavenging of (DPPH) was assayed by the modified method ¹⁴, (results were expressed as AEAC (ascorbic acid equivalent antioxidant capacity) in grams and the calculation was done according to the followed equation).¹⁴

$$AEAC = (\mu\text{g AA/g}) = IC50 (AA)/IC50 (\text{sample}) \times 1\text{g}$$

3. RESULTS AND DISCUSSION

3.1. Isolation & phytochemical screening

Ethyl acetate fraction was carefully injected to silica gel column chromatography and the elution was done by (CH₂Cl₂: MeOH) with different concentrations (100:0→60:40) then the collective Fractions was subjected to successive column chromatography on Sephadex LH-20 using methanol to offered four compounds which all subjected to ESI–mass spectroscopy (Ain shams University), and 1H- and 13C-NMR (Cairo university) measurements spectrometer operation at 400 MHz (for 1H) and 100 MHz (for 13C). The phytochemical screening of the 70% alcohol extract of Malpighia glabra. L leaves are presented in Table 1.

Table 1: Phytochemical screening results of the 70% methanolic extract of *Malpighia glabra. L* leaves*

1-	Crystalline sublimate	- ve
2-	Volatile oil	+ ve
3-	Carbohydrates and/or glycosides	+ ve
4-	Oxidase enzyme	- ve
	Flavonoids:	++ ve
5-	- Aglycones	++ ve
	- Glycoside	+ ve
6-	Tannins	++ ve
7-	Sterols and/or triterpenes	++ ve
8-	Saponins	- ve
9-	Cardiac glycosides	- ve
10-	Alkaloids	- ve
11-	Anthraquinone derivatives	- ve

*- ve = Absent + ve = Present ++ve = Strongly present

Screening of Malpighia glabra.L leaves phytochemical substances revealed the existence of volatile oil, carbohydrates and/or glycosides, sterols, tanins and/or triterpenes and flavonoids, and absence of crystalline sublimate, cardiac glycosides, saponins, alkaloids, oxidase enzyme and Anthraquinone derivatives.

3.2. Isolated compounds

Chromatographic investigation of Malpighia glabra.L leaves extract yielded four compounds, compound 1, Caffeic acid(figure.1), compound 2, Chlorogenic acid(figure. 2), compound 3, Quercetin (figure.3), & compound 4, Kaempferol (figure.4)

Compound 1: It is appeared as white powder with amorphous character. ESI-MS of the compound

was measured on (TQD triple quadruple instrument (Waters Corporation, Milford, instrument), and showed $[M-H]^+$ peak at m/z 179.1, which is compatible with the molecular formula $C_9H_8O_4$. The 1H NMR spectrum of the compound exhibited signals which is characteristic for δ 7.57 (1H, d, $J = 16$ Hz, H-7), 7.06 (1H, d, $J = 2.0$ Hz, H-2), 6.96 (1H, dd, $J = 8.0, 2.0$ Hz, H-6), 6.94 (1H, d, $J = 8.0$ Hz, H-5), 6.26 (1H, d, $J = 16$ Hz, H-8).

The ^{13}C -NMR spectral data of the compound showed signals at δ 169.80 (C-9), 148.03 (C-4), 145.76 (C-3), 145.5C-7), 126.43 (C-1), 121.57 (C-6), 115.15 (C-5), 114.11 (C-2), 113.77 (C-8). From these data, compound 1 was identified as caffeic acid, which is confirmed by the direct comparison with published data ¹⁵.

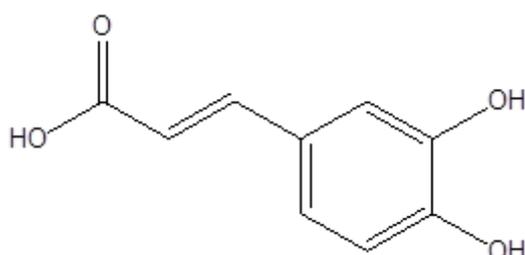


Figure 1: Chemical structure of Caffeic acid ¹⁵

Compound 2: It was isolated as white powder with amorphous shape. ESI-MS of the compound revealed $[M-H]^+$ peak at m/z 353.4, which is compatible with the molecular formula $C_{16}H_{18}O_9$.

The 1H NMR spectrum of the compound exhibited signals which characteristic for δ 7.44 (1H, d, $J = 16$ Hz, H-7'), 7.04 (1H, d, $J = 2.0$ Hz, H-2'), 6.99 (1H, dd, $J = 8.0, 2.0$ Hz, H-6'), 6.78 (1H, d, $J = 8.0$ Hz, H-5'), 6.17 (1H, d, $J = 16$ Hz, H-8'), 5.08 (1H, ddd, $J = 6.8, 6.8, 4.4$ Hz, H-5), 3.95 (1H, ddd, $J = 7.2, 3.6, 3.6$ Hz, H-3), 3.58 (1H, dd, $J = 6.8, 2.4$ Hz, H-4), 2.04 (1H, m, H-2ax), 2.01 (1H, m, H-6ax), 1.96 (1H, dd, $J = 13.2, 4.0$ Hz, H-6eq), 1.82 (1H, dd, $J = 13.2, 7.6$ Hz, H-2eq)

The ^{13}C -NMR spectrum of the compound revealed signals at δ 175.00 (C-7), 165.83 (C-9'), 148.40 (C-4'), 145.63 (C-3'), 145.03 (C-7'), 125.70 (C-1'), 121.46 (C-6'), 115.85 (C-5'), 114.84 (C-2'), 114.40 (C-8'), 73.59 (C-1), 70.95 (C-5), 70.51 (C-4), 68.20 (C-3), 37.28 (C-2), 36.39 (C-6).

From the data mentioned above, compound 2 was identified as chlorogenic acid, which confirmed by direct comparison with published data ¹⁶

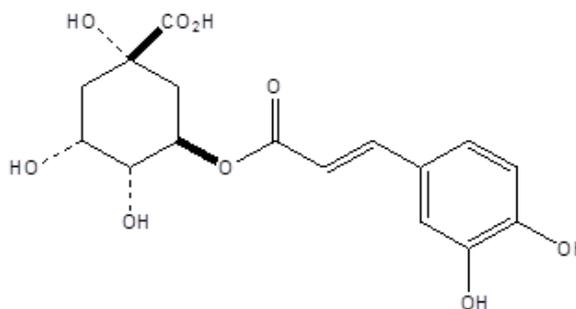


Figure 2: Chemical structure of Chlorogenic acid.¹⁶

Compound 3: The compound was isolated as a yellow powder with amorphous characters. The ESI-MS of the compound showed a peak at m/z 301.0 $[M-H]^+$ indicating that the molecular formula is $C_{15}H_{10}O$. The 1H -NMR spectrum of the compound showed a clear flavonol type pattern of aromatic proton signals; two meta coupled protons each at δ 6.19 ppm (d, $J = 2.0$, H-6) and δ 6.41 ppm (d, $J = 2.0$, H-8), doublet of one proton at δ 6.90 ppm (d, $J = 8.4$ Hz, H-5') and a meta coupled doublet of one proton at δ 7.68 ppm (d, $J = 2.0$, H-2') and H-6' appear as doublet of doublet at 7.55 ppm (dd, $J = 8.4, 2.0$, H-6'). This data indicated that the compound is a 3, 5, 7, 3, 4'-penta oxygenated flavone derivative, which is in a good agreement with Quercetin. The ^{13}C -NMR (100 MHz, DMSO- d_6) spectrum of the compound supporting the structure assignment made above, revealing the presence of 15 carbon signals, including 8 oxygenated at [δ 145.11 (C-2), 160.78 (C-5), 163.97 (C-7), 156.21 (C-9), 146.86 (C-3'), 135.78 (C-3), δ 175.89 (C-4) and 147.75 (C-4')]. 7 non-oxygenated at [δ 98.26 (C-6), 93.42 (C-8), 103.07 (C-10), 122.04 (C-1'), 115.14 (C-2'), 115.67 (C-5') and 120.05 (C-6')] ^{17,18}. Compound 3 is Quercetin according to the literature data. ^{17,18}

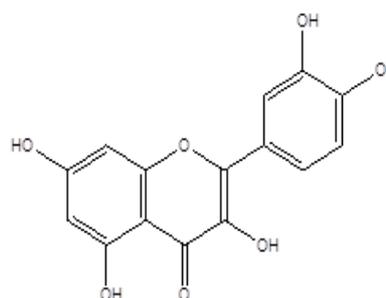


Figure 3: Chemical structure of Quercetin ^{17,18}

Compound 4: Compound 4 was isolated as fine yellow needles. The ESI-MS spectrum of compound 4, showed a molecular ion peak at m/z 286 $[M]^+$, corresponded to the molecular formula $C_{15}H_{10}O_6$.

Peaks due to RDA fragmentation at m/z 153 corresponding to ring-A, respectively, indicating a kaempferol nucleus

The ¹H-NMR spectrum of compound 4 revealed a clear Flavonol-type pattern of aromatic proton signals; two meta-coupled protons each at δ 6.19 ppm (d, j=2 H-6) and δ 6.44ppm (d, j =2 H-8) and an (AA'BB') system, characteristic to a 1,4-disubstituted aromatic B ring at δ 8.06 and 6.94 (each 2H, each d, J= 8.8 Hz, H-2', 6' and H-3', 5', respectively).

The ¹³C-NMR spectrum of compound 4 showed 15 carbon signals at 147.24 (C- 2), 136.12(C- 3), 176.36(C- 4), 161.17(C- 5), 98.71(C- 6), 164.52(C- 7), 93.96(C- 8), 156.65(C- 9), 103.45(C- 10), 122.14(C- 1'), 129.96(C- 2'/6'), 115.91(C- 3'5'), 159.66(C- 4') identical with those reported for kaempferol nucleus. The p-substitution of B ring was confirming by the same chemical shifts for the equivalent C-3'/C-5' and C-2'/C-6'. The ¹H-NMR & ¹³C-NMR spectral data were similar to with those reported for kaempferol ^{17,18}. Compound 4 is kaempferol according to the literature data. ^{17,18}.

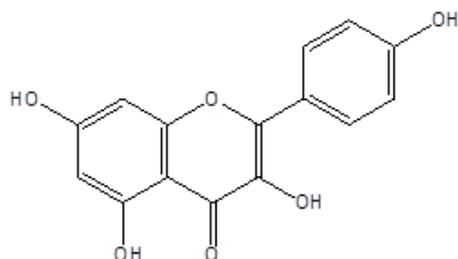


Figure 4: Chemical structure of Kaempferol ^{17,18}

The total phenolics of *Malpighia glabra*.L leaves extract was determined spectrophotometrically at 765 nm which is found to be 96.36 mg GAE/g of extract compared to gallic acid as in graph (figure.5)

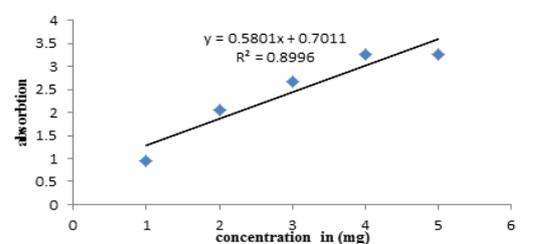


Figure 5: Standard curve for Gallic acid ¹²

3.4. Quantification of total flavonoid contents:

The results that observed in the graph (fig.6) had declared that *Malpighia glabra*. L rich in flavonoids (total flavonoid content: 30.71 mg QE/g extract).

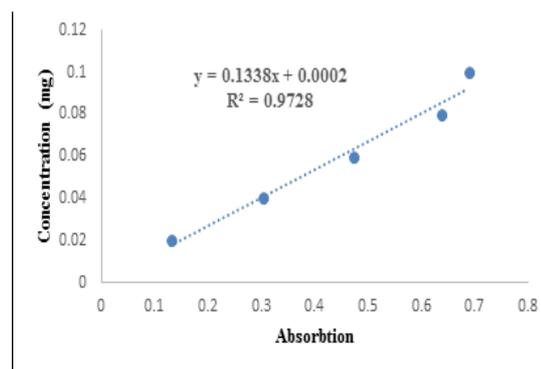


Figure 6: Standard curve for quercetin ¹³

Phenolic acids, flavonoids content and free radical scavenging agents ^{19,20} have a positive relation to each other. Concentration of phenolic compounds and the antioxidant effect have a significant linear correlation, many studies had ensured the correlation between the phenolics and free radical scavenging activity.

3.5. Antioxidant assay

Plant extract which contains high levels of phenolics shows great antioxidant activity and resulted in free radical scavenging activity ²¹

Complementing this result, DPPH test of *Malpighia glabra*. L leaves extract shows significant decrement of DPPH radicals which is nearly similar to the considerable antioxidant agent “ascorbic acid”, and lead to DNA protection specially in fruits that contains antioxidant compounds²², carotenoids and poly phenols ²³⁻²⁵ but consistency of vit C in unripe fruits is higher than ripe fruits.²⁶

The DPPH assay result of *Malpighia glabra*. L using ascorbic acid as positive control shows that (IC₅₀ = 5.159 µg/ml) validate the assay, while the amount of the extract which required to capture the DPPH radicals is (IC₅₀ = 4.98 µg/ml) acerola leaves (Table 2).

Table 2: Antioxidant assay of **Malpighia glabra. L** extract

sample	Inhibition per concentration		IC ₅₀ (mg/ml)	IC ₅₀ (µg/ml)	AEAC(µg/ml)
	100mg/ml	gm/ml			
ascorbic acid	100.4	1000.4	51.59	5.16	1.000
extract	96.9	969	49.8	4.98	0.965

4. CONCLUSIONS

Separation of four pure compounds from Ethyl acetate fraction of *Malpighia glabra*.L leaves, that compounds are Caffeic acid, Chlorogenic acid, Quercetin & Kaempferol. Moreover, **Malpighia glabra. L** extract shows high values of total phenolic and total flavonoid contents reflecting high free radicals scavenging activity.

Conflict of interest: All authors declared no conflict of interest.

Author contribution: AMF performed the extraction, participated in chromatographic separation of the pure compounds and wrote the paper. AIM and MAE revised the paper and conceived the phytochemical and biological study. WAE shared in the biological study. MSA participated in the structure elucidation of the isolated compounds, wrote and conceived the study.

Acknowledgements: A sincerely thanks to the Faculty of Pharmacy (Girls), Al-Azhar University for the providing us with the laboratory facilities which the work could not be accomplished without it

Funding: There is no funding source for this study.

REFERENCES

1. Lin L, Liu YC, Huang JL, Liu XB, Qing ZX, Zeng JG, et al. Medicinal plants of the genus *Macleaya* (*Macleaya cordata*, *Macleaya microcarpa*): A review of their phytochemistry, pharmacology, and toxicology. 2018;32(1):19-48.
2. Pandey N, Meena RP, Rai KS, Rai SP. Medicinal Plants Derived Nutraceuticals : A re-emerging health aid. 2011; 2(4): 419-441. (Review).

3. Delva L, Schneider RGJFRI. Acerola (*Malpighia emarginata* DC): production, postharvest handling, nutrition, and biological activity. 2013;29(2):107-26.
4. de Assis SA, Fernandes FP, Martins ABG, de Faria Oliveira OMMJF. Acerola: importance, culture conditions, production and biochemical aspects. 2008;63(2):93-101.
5. Prakash A, Baskaran R. Acerola, an untapped functional superfruit: a review on latest frontiers. J Food Sci Technol. 2018;55(9):3373-84.
6. da Silva Nunes R, Kahl VFS, da Silva Sarmiento M, Richter MF, Costa-Lotufo LV, Rodrigues FAR, et al. Antigenotoxicity and antioxidant activity of Acerola fruit (*Malpighia glabra* L.) at two stages of ripeness. 2011;66(2):129-35.
7. Belwal T, Devkota HP, Hassan HA, Ahluwalia S, Ramadan MF, Mocan A, et al. Phytopharmacology of Acerola (*Malpighia* spp.) and its potential as functional food. 2018;74:99-106.
8. Munier Rjbdlscdf . * Bases Azotees (Alcaloids, Amines, Vitamins) et Acides Organiques Hydrosolubles. 1952;19(9-10):852-73.
9. Gorin PA, Mazurek MJJoC. Further studies on the assignment of signals in ¹³C magnetic resonance spectra of aldoses and derived methyl glycosides. 1975;53(8):1212-23.

10. de Bruyne T, Pieters LA, Dommisse RA, Kolodziej H, Wray V, Domke T, et al. Unambiguous assignments for free dimeric proanthocyanidin phenols from 2D NMR. 1996;43(1):265-72.
11. Bae Y-s, Burger JF, Steynberg JP, Ferreira D, Hemingway RWJP. Flavan and procyanidin glycosides from the bark of blackjack oak. 1994;35(2):473-8.
12. Singleton VL, Orthofer R, Lamuela-Raventós RMJMie. [14] Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. 1999;299:152-78.
13. Zhishen J, Mengcheng T, Jianming WJFc. The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. 1999;64(4):555-9.
14. Yamaguchi T, Takamura H, Matoba T, Terao JJB, biotechnology, biochemistry. HPLC method for evaluation of the free radical-scavenging activity of foods by using 1, 1-diphenyl-2-picrylhydrazyl. 1998;62(6):1201-4.
15. Tošović JJKJoS. Spectroscopic features of caffeic acid: theoretical study. 2017(39):99-108
16. Agrawal Pkjsioc. Carbon-13 NMR of flavonoids. 1989;39:XVI-564.
17. Markham K, Ternai B, Stanley R, Geiger H, Mabry TJT. Carbon-13 NMR studies of flavonoids—III: Naturally occurring flavonoid glycosides and their acylated derivatives. 1978;34(9):1389-97.
18. Oki T, Masuda M, Furuta S, Nishiba Y, Terahara N, Suda IJJoFS. Involvement of anthocyanins and other phenolic compounds in radical-scavenging activity of purple-fleshed sweet potato cultivars. 2002;67(5):1752-6.
19. Duan X, Wu G, Jiang YJM. Evaluation of the antioxidant properties of litchi fruit phenolics in relation to pericarp browning prevention. 2007;12(4):759-71.
20. Tosun M, Ercisli S, Sengul M, Ozer H, Polat T, Ozturk EJBR. Antioxidant properties and total phenolic content of eight Salvia species from Turkey. 2009;42(2):175-81.
21. Katalinic V, Milos M, Kulisic T, Jukic MJFc. Screening of 70 medicinal plant extracts for antioxidant capacity and total phenols. 2006;94(4):550-7.
22. Franke SIR, Prá D, da Silva J, Erdtmann B, Henriques JAPJMRGT, Mutagenesis E. Possible repair action of Vitamin C on DNA damage induced by methyl methanesulfonate, cyclophosphamide, FeSO₄ and CuSO₄ in mouse blood cells in vivo. 2005;583(1):75-84.
23. Leong L, Shui GJFc. An investigation of antioxidant capacity of fruits in Singapore markets. 2002;76(1):69-75.
24. Paredes-López O, Cervantes-Ceja ML, Vigna-Pérez M, Hernández-Pérez TJPffhn. Berries: improving human health and healthy aging, and promoting quality life—a review. 2010;65(3):299-308.
25. Seeram NP, Aviram M, Zhang Y, Henning SM, Feng L, Dreher M, et al. Comparison of antioxidant potency of commonly consumed polyphenol-rich beverages in the United States. 2008;56(4):1415-22.
26. Jawaheer B, Goburdhun D, Ruggoo AJPFfHN. Effect of processing and storage of guava into jam and juice on the ascorbic acid content. 2003;58(3):1-12.