

Novel Mannich Bases Derived from 2-Substituted Benzimidazole and (Thio)Hydantoin Moieties as Potent Histone Deacetylase 6 (HDAC6) Inhibitors

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Abstract: Novel Mannich bases derived from 2-substituted benzimidazole and (thio) hydantoin moieties, were synthesized as histone deacetylase 6 (HDAC6) inhibitors with potential cytotoxic activity. All derivatives were tested *In vitro* against HDAC6 enzyme. IR, ¹H NMR, ¹³C NMR and mass spectroscopy confirmed the structure of synthesized compounds. All tested compounds significantly inhibited HDAC6 at nanomolar level. Compound **2c** was the most potent presenting significant HDAC6 inhibitory activity (IC₅₀ = 97.35 ± 5.7 nM), nearly equipotent to SAHA reference drug (IC₅₀ = 91.73 ± 5.4 nM). *In vitro* cytotoxicity study was also carried out. Compound **2c** exhibited potent cytotoxic activity against the tested cells (CCRF-CEM and MOLT-4) showing one digit micromolar IC₅₀s. Compound **2c** (IC₅₀ = 3.66 ± 0.22 μM) was 2-fold more active than SAHA reference drug (IC₅₀ = 6.8 ± 0.41 μM) against MOLT-4 cell line. Furthermore, a docking study demonstrated the ability of target Mannich bases to achieve an excellent fitting inside the binding site of HDAC6 enzyme.

Keywords: benzimidazole; (thio)hydantoin; HDAC6; Mannich bases; cytotoxicity.

Article history: Received 2021-06-01

Revised 2021-07-04

Accepted 2021-08-03

1. INTRODUCTION

Leukemia is a lethal type of cancer that leads to hematopoietic malignancies with relatively poor (20-40%) patients' survival. Even though the exploration for various therapy, leukemia still the sixth leading cause for death in USA¹. Recently, a lot of work has been directed to epigenetic changes that accompanied the development of hematopoietic malignancies². These changes play essential roles in regulating gene expression that can eventually induce oncogenic neoplasms³. Among these epigenetic changes is the modification of histone acetylation which can account for alteration of DNA accessibility and chromatin structure⁴. Histone acetylation is regulated by histone acetyltransferase (HAT) and histone deacetylase (HDAC) enzymes⁵.

Among HDAC family, HDAC6 is a unique Zn²⁺-dependent enzyme which plays a vital role in microtubule dynamics and so affects cell proliferation and survival. In addition, HDAC6 acts on nonhistone

substrates eg. Hsp90 and peroxiredoxin. HDAC6 has a direct effect on cellular activities including motility, growth of cells, migrations and adhesion⁶. Furthermore, HDAC6 is highly expressed during the development of tumors^{7,8}. It has also been reported that inhibition of HDAC6 resulted in apoptosis. Therefore, HDAC6 has been considered as an attractive target for designing potent anticancer candidates. Recently, several pan-HDAC inhibitors eg SAHA (suberoylanilide hydroxamic acid), were approved by FDA (Food and Drug Administration) for the management of hematological cancers^{9,10}. HDAC6 inhibitors eg. tubastatin A¹¹ and citarinostat (ACY-241)¹² have also been reported. Structurally, HDAC inhibitors have common pharmacophoric features: cap-linker-zinc binding group (ZBG) (**Figure.1**).

Benzimidazole scaffold has attracted considerable attention for the discovery and development of antitumor candidates^{13,14}. Several benzimidazole-containing drugs such as nocodazole and veliparp

have been clinically approved for cancer treatment¹⁵. Moreover, the HDAC inhibitory activity of benzimidazole derivatives has been reported by several research groups¹⁶ (**Figure.2**).

On the other hand, hydantoin and thiohydantoin derivatives are five membered heterocyclic compounds that attracted much attention in medicinal chemistry. Hydantoin-based compounds have been reported to exhibit considerable anticancer activities¹⁷. On the other hand, owing to their chelating properties, hydantoin and thiohydantoin have also been recognized to inhibit different metalloenzymes like matrix metalloproteinase¹⁸ and tryosinase¹⁹. For these observations, we assumed that hydantoin and thiohydantoin moieties might be considered as non-hydroxamate zinc chelating groups essential for HDAC inhibitory activity.

In the present study, novel Mannich bases derived from 2-substituted benzimidazole and (thio)hydantoin moieties were developed as HDAC6 inhibitors (**Figure.3**). Following the basic requirements for designing HDAC inhibitors, the benzimidazole nucleus is utilized as a capping moiety that interacts with the enzyme external surface. Whereas, the (thio) hydantoin moiety represents non-hydroxamate ZBG that can chelate Zn²⁺ metal at HDAC enzyme binding site. The benzimidazole and the (thio) hydantoin moieties were separated by a CH₂ linker. Different substituents (R= H, CH₃, Ph) were also added to the 2-position of benzimidazole ring to discover substituent effect on HDAC inhibition. In this work, all derivatives were screened against HDAC6. In vitro cytotoxicity and docking study were also conducted.

2. METHODS

2.1. Chemistry

For general remarks, see supplementary file. The starting compounds **1a-c** were prepared as illustrated in the literature reviews^{20, 21}.

Synthesis of benzimidazole derivatives (**1a-c**)

A mixture of *o*-phenylenediamine (0.1 mol) and the corresponding acids, formic acid, acetic acid and benzoic acid (0.1 mol) in 4 N HCl (15 ml), was refluxed for 6-8 h. The reaction mixture was neutralized with 10% sodium hydroxide solution. The precipitated product was filtered, dried and crystallized from absolute ethanol. **Yield** 80-90%.

Synthesis of Mannich bases **2a-f**:

A new series of Mannich bases were prepared according to reported procedures²²⁻²⁵. A mixture of benzimidazole derivatives **1a-c** (2 mmol), and 38% formaldehyde solution (2 mmol) was added into the

solution of hydantoin or thiohydantoin (2 mmol) in absolute ethanol (10 ml). The reaction mixture was refluxed for 3-4 h. The precipitated products were filtered, dried and crystallized from 99% ethanol.

5-((1*H*-benzo[*d*]imidazol-1-yl)methyl)imidazolidine-2,4-dione (**2a**)

Yield:(73.90%); **m.p.** 190- 191 °C; **IR (KBr)** (cm⁻¹): 3220, 3190 (2NH), 1780, 1698 (2C=O); **¹H NMR** (DMSO-*d*₆) δ (ppm): 3.97 (d, 1H, CH₂, *J* = 3.9 Hz), 4.07(d, 1H, CH₂, *J* = 3.9 Hz), 4.72- 4.76 (m, 1H, CH), 6.28 (s, 1H, NH, D₂O exchangeable), 6.42 (s, 1H, NH, D₂O exchangeable), 7.20-7.31 (m, 2H, benzimidazole-H), 7.72 (d, 1H, benzimidazole-H, *J* = 8 Hz), 8.21 (d, 1H, benzimidazole-H, *J* = 7.6 Hz), 8.33 (s, 1H, benzimidazole-H); **¹³C NMR** (DMSO-*d*₆) δ (ppm): 48.25 (CH₂), 60.79 (CH), 111.07, 119.97, 123.29, 123.32, 133.74, 144.70, 158.81(N-C=N), 169.84, 174.36 (2C=O); **MS *m/z* (%)**: 230.11 (M⁺, 8.17), 132.69 (1.37), 131.18 (2.27), 100.21(100); **Analysis** % for C₁₁H₁₀N₄O₂ (230) Calcd. (Found) C, 57.39 (57.13), H, 4.38 (4.09), N, 24.34 (23.99).

5-((2-methyl-1*H*-benzo[*d*]imidazol-1-yl)methyl)imidazolidine-2,4-dione (**2b**)

Yield:(60.93%); **m.p.** 90- 92 °C; **IR (KBr)** (cm⁻¹): 3317, 3300 (2NH), 1774, 1693 (2C=O); **¹H NMR** (DMSO-*d*₆) δ (ppm): 2.60 (s, 3H, CH₃) 4.01- 4.11 (m, 2H, CH₂), 4.56-4.77 (m, 1H, CH), 7.16-7.24 (m, 2H, benzimidazole-H), 7.54-7.58 (m, 2H, benzimidazole-H), 8.11 (s, 1H, NH, D₂O exchangeable), 10.92 (s, 1H, NH, D₂O exchangeable); **¹³C NMR** (DMSO-*d*₆) δ (ppm): 13.65 (CH₃), 47.70 (CH₂), 61.39 (CH), 110.47, 118.62, 122.62, 122.9, 134.86, 142.2, 158.83 (N-C=N), 169.84, 172.36 (2C=O); **MS *m/z* (%)**: 244.22 (M⁺, 9.01), 241.62 (1.32), 130.42 (100); **Analysis** % for C₁₂H₁₂N₄O₂ (244) Calcd. (Found) C, 59.01 (58.81), H, 4.95 (4.63), N, 22.94 (23.14).

5-((2-phenyl-1*H*-benzo[*d*]imidazol-1-yl)methyl)imidazolidine-2,4-dione (**2c**)

Yield:(75.60%); **m.p.** 280- 282 °C; **IR (KBr)** (cm⁻¹): 3209 (2NH), 1766, 1693 (2C=O); **¹H NMR** (DMSO-*d*₆) δ (ppm): 4.00-4.06 (m, 2H, CH₂), 4.73-4.77 (m, 1H, CH), 7.27-7.31 (m, 2H, benzimidazole-H), 7.49-7.54 (t, 1H, phenyl-H), 7.56 (s, 1H, NH, D₂O exchangeable), 7.57 (s, 1H, NH, D₂O exchangeable), 7.58-7.69 (m, 2H, phenyl-H), 7.98 (d, 2H, benzimidazole-H, *J* = 8.4 Hz), 8.24-8.27 (d, 2H, phenyl-H, *J* = 8 Hz); **¹³C NMR** (DMSO-*d*₆) δ (ppm): 50.55 (CH₂), 60.27 (CH), 115.34, 123.42, 127.30, 129.04, 129.10, 129.20, 129.28, 129.57, 129.75, 131.08, 131.23, 138.20, 151.15 (N-C=N), 167.82, 172.36 (2C=O); **MS *m/z* (%)**: 306.11 (M⁺, 17.72), 268.82 (1.42), 64.25 (100); **Analysis** % for C₁₇H₁₄N₄O₂ (306) Calcd. (Found) C, 66.66 (67.05), H, 4.61 (4.39), N, 18.29 (18.59).

5-((1*H*-benzo[*d*]imidazol-1-yl)methyl)-2-thioxoimidazolidin-4-one (2d)

Yield:(80.50%); **m.p.** 108- 110 °C; **IR (KBr)** (cm⁻¹): 3143, 3101 (2NH), 1755, 1616 (2C=O), 1199 (C=S); **¹H NMR** (DMSO-*d*₆) δ (ppm): 3.38-3.46 (m, 2H, CH₂), 4.55- 4.99 (m, 1H, CH), 6.78 (s, 1H, NH, D₂O exchangeable), 7.22-7.31 (m, 2H, benzimidazole-H), 7.69 (d, 2H, benzimidazole-H, *J* = 8 Hz), 8.29 (s, 1H, benzimidazole-H), 10.01 (s, 1H, NH, D₂O exchangeable); **¹³C NMR** (DMSO-*d*₆) δ (ppm): 50.58 (CH₂), 67.80 (CH), 111.38, 119.82, 122.25, 122.89, 133.75, 144.11, 144.34 (N-C=N),174.36 (C=O), 181.78 (C=S); **MS *m/z* (%)**: 246.46 (M⁺, 12.94), 244.21 (4), 240.47 (4.40), 64.40(100); **Analysis %** for C₁₁H₁₀N₄OS (246) Calcd. (Found) C, 53.64 (53.98), H, 4.09 (3.80), N, 22.75 (23.06).

5-((2-methyl-1*H*-benzo[*d*]imidazol-1-yl)methyl)-2-thioxoimidazolidin-4-one (2e)

Yield:(69.45%); **m.p.** 139-140 °C; **IR (KBr)** (cm⁻¹): 3120 (2NH), 1755, 1616 (2C=O), 1149 (C=S); **¹H NMR** (DMSO-*d*₆) δ (ppm): 2.56 (s, 3H, CH₃) 3.61-3.82 (m, 2H, CH₂), 4.82-4.99 (m, 1H, CH), 6.66 (s, 1H, NH, D₂O exchangeable), 7.16-7.24 (m, 2H, benzimidazole-H), 7.53-7.57 (m, 2H, benzimidazole-H_{4,7}), 10.01 (s, H, NH, D₂O exchangeable); **¹³C NMR** (DMSO-*d*₆) δ (ppm): 13.74 (CH₃), 60.27 (CH₂), 66.35 (CH), 110.74, 118.32, 122.14, 122.65, 134.99, 142.02, 152.25 (N-C=N),174.16 (C=O), 181.43 (C=S); **MS *m/z* (%)**: 260.41(M⁺, 3.32), 257.29 (3.66), 41.27 (100); **Analysis %** for C₁₂H₁₂N₄OS (260) Calcd. (Found) C, 55.37 (55.69), H, 4.65 (5.03), N, 21.52 (21.14).

5-((2-phenyl-1*H*-benzo[*d*]imidazol-1-yl)methyl)-2-thioxoimidazolidin-4-one (2f)

Yield:(59.30%); **m.p.** 158- 160 °C; **IR (KBr)** (cm⁻¹): 3232, 3217 (2NH), 1755, 1681 (2C=O), 1180 (C=S); **¹H NMR** (DMSO-*d*₆) δ (ppm): 3.56-3.83 (m, 2H, CH₂), 4.83- 4.97 (m, 1H, CH), 6.37 (s, 1H, NH, D₂O exchangeable), 7.27-7.30 (m, 2H, benzimidazole-H), 7.49-7.68 (m, 3H, phenyl-H), 7.97 (d, 2H, benzimidazole-H, *J* = 7.4 Hz), 8.24 (d, 2H, phenyl-H, *J* = 7 Hz), 10.01 (s, 1H, NH, D₂O exchangeable); **¹³C NMR** (DMSO-*d*₆) δ (ppm): 55.20 (CH₂), 69.60 (CH), 115.14, 124.08, 127.58, 127.88, 129.04, 129.34, 129.69, 129.73,131.20, 131.69, 133.34, 136.88, 150.68 (N-C=N), 167.79 (C=O), 173.90 (C=S); **MS *m/z* (%)**: 322.88 (M⁺, 6.11), 318.77(5.11), 59.15 (100); **Analysis %** for C₁₇H₁₄N₄OS (322) Calcd. (Found) C, 63.34 (62.98), H, 4.38 (4.75), N, 17.38 (17.67).

2.1.2. In vitro HDAC6 inhibition assay:

HDAC6 inhibitory activity was measured using BioVision's HDAC6 inhibitor screening kit using SAHA as a reference drug²⁶. For details, see supplementary file.

2.1.3. Anticancer activity:

The anticancer activity of compound **2c** against CCRF-CEM and MOLT-4 cancer cells was estimated using MTT-based assay kit, Sigma²⁷⁻²⁹. For details, see supplementary file.

2.2. Figures, Tables and Schemes

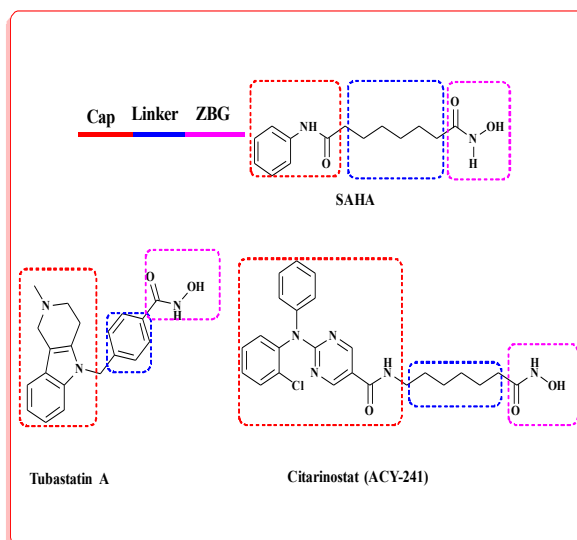


Figure 1. reported histone deacetylase inhibitors

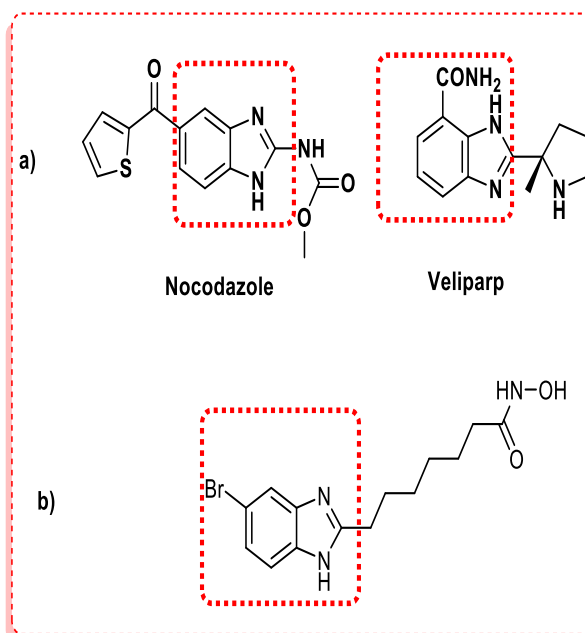


Figure2. a) Clinically approved enzimidazole -containing anticancer drugs.

b) Reported benzimidazole as HDAC inhibitors.

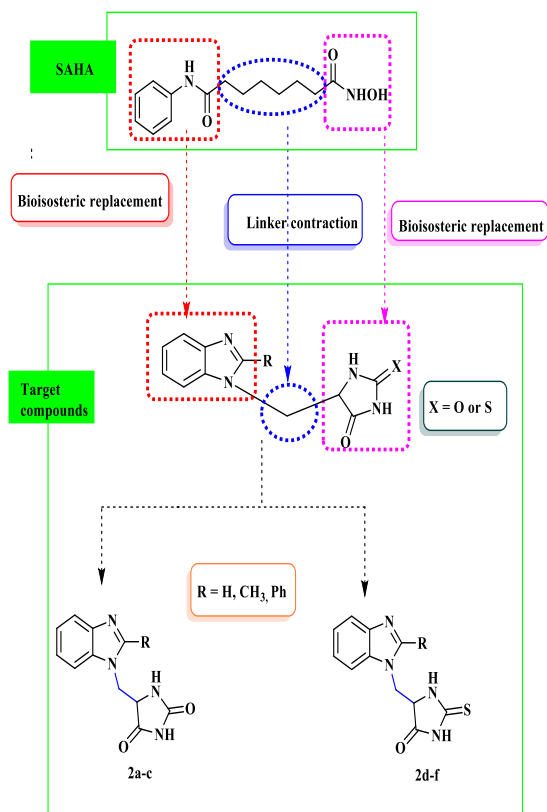


Figure 3. Design of target compounds as HDAC6 inhibitors.

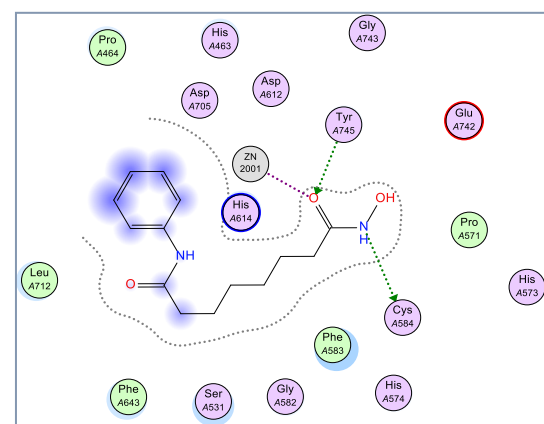
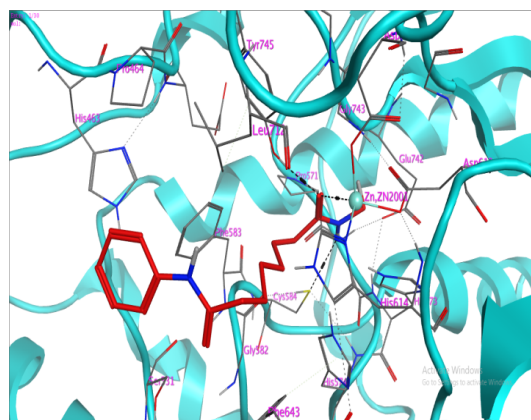


Figure 4a. 2D and 3D of SAHA docked into the active site of HDAC6.

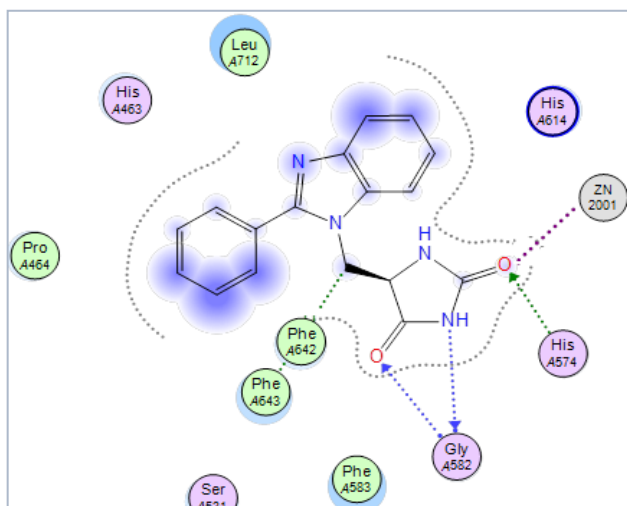
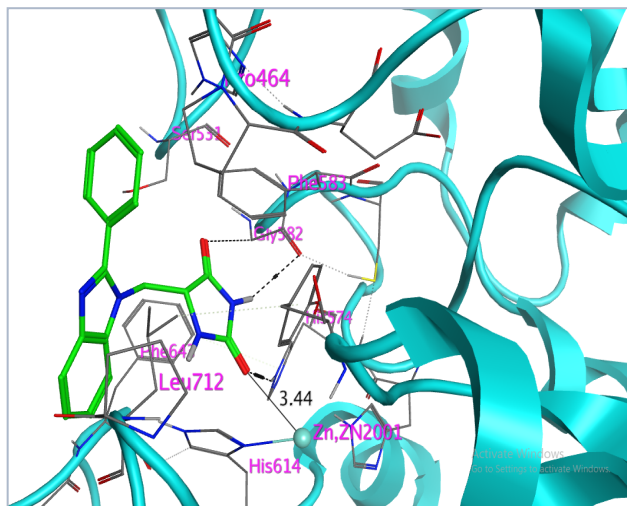


Figure 4b. 2D and 3D of compound 2c docked into the active site of HDAC6.

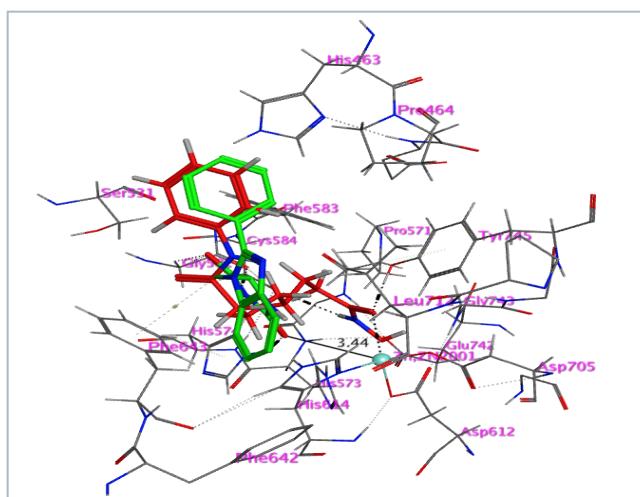


Figure 4c. The overlay of docked 2c (green color) and SAHA (red color) within the active site of HDAC6.

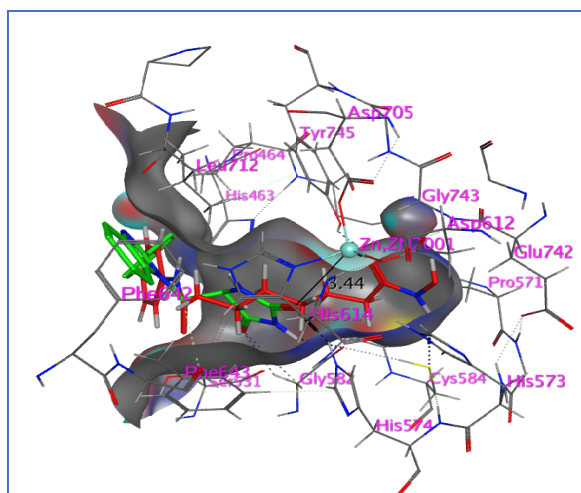


Figure 4d. Side view for compound 2c (green colour) and SAHA (red colour) inside the binding pocket of HDAC6.

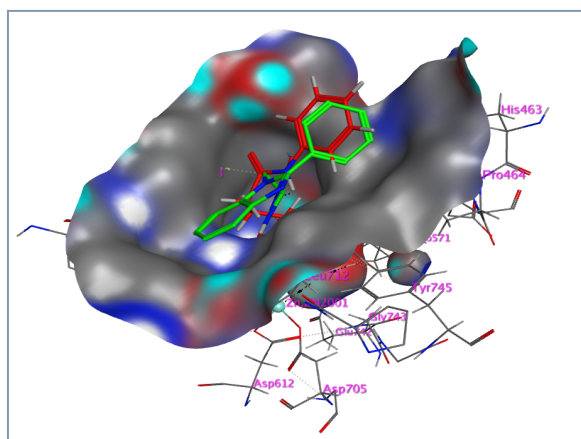
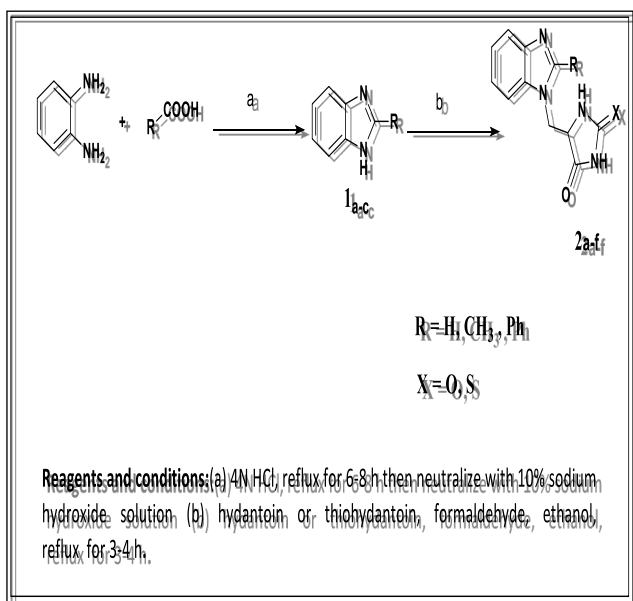


Figure 4e. Top view for compound 2c (green colour) and SAHA (red colour) inside the binding pocket of HDAC6



Scheme 1: Synthesis of target Mannich bases 2a-f.

Table 1: HDAC6 inhibitory activity of target compounds.

Comp. No.	Structure	IC ₅₀ (nM)
2a		279.59± 16
2b		166.09± 9.7
2c		97.35± 5.7
2d		209.27± 12
2e		386.38± 23
2f		179.73± 11

SAHA		91.73±5.44
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Table 2: *In vitro* anticancer activity against two human leukaemia cancer cell lines, CCRF-CEM and MOLT-4

Comp. No.	Cytotoxicity	
	CCRF-CEM	MOLT-4
2c	9.67±0.58	3.66±0.22
SAHA	3.83±0.23	6.8±0.41

Table 3: Docking results for binding interaction of SAHA and 2c in HDAC6 binding site.

Comp. No	Docking score (Kcal/mol)	No. of H bonds	Amino acid residues (bond length Å)	Atom of compound			
SAHA	- 6.64	2	Cys 584 (4.07)	NH of hydroxymate			
			Tyr 745 (2.65)	C=O of hydroxymate			
			Zn (2.63)	O of C=O of hydroxymate			
			Leu 712				
			Ser 531	Phenyl capping group			
			Phe 583				
			Phe 643				
			His 614				
			Pro 464				
			2c	-4.68	3	His574 (2.89)	C=O of hydantoin
						Gly 582 (3.23)	C=O of hydantoin
Gly 582 (3.36)	NH of hydantoin						
Phe 643 (4.45)	CH ₂ of the linker moiety						
Zn (3.44)	O of hydantoin						
Leu 712	2-phenyl						
Ser 531	benzimidazole						
Phe 583	capping group						
Phe 642							
Phe 643							
His 614							
His 463							
Pro 464							

3. RESULTS

3.1 Chemistry

In the present work, 2-substituted-benzimidazoles **1a-c** were synthesized via the

reaction of *o*-phenylenediamine with the appropriate carboxylic acid following reported procedure^{20,21} (**Scheme 1**).

It is worth noting that Mannich reaction has a great application in medicinal chemistry as Mannich bases have interesting biological activities eg. anticancer, antibacterial, antiviral and anti-inflammatory²². Additionally, aminomethylation of drugs may improve their delivery in the human body and increase the hydrophilic properties through addition of a polar functional group²². Following reported procedures²³⁻²⁵, benzimidazoles **1a-c** were reacted with hydantoin or thiohydantoin and formaldehyde solution to give Mannich bases **2a-f** through aminoalkylation reaction (**Scheme 1**).

3.2 HDAC6 assay:

Mannich bases **2a-f**, were evaluated against HDAC6 using SAHA as a reference standard (**Table 1**). All derivatives efficiently suppressed HDAC6 activity showing IC₅₀ values in the nanomolar range (97.356 to 368.38 nM), relative to SAHA reference drug (IC₅₀ = 91.732 ± 5.4 nM). Compound **2c** was the most active against HDAC6 enzyme showing a significant HDAC6 inhibitory activity (IC₅₀ = 97.356 ± 5.7 nM), nearly equipotent to SAHA (IC₅₀ = 91.732 ± 5.4 nM). Analogues **2b**, **2d** and **2f** revealed half potency of SAHA. While, compounds **2a** and **2e** were the least potent against HDAC6 enzyme.

In vitro anticancer activity:

As mentioned before, SAHA was approved by FDA for the treatment of various hematological cancers including acute myeloid leukemia and multiple myeloma, thus its effectiveness against leukemia is mainly assigned to HDAC inhibition. Thus, compound **2c**; the most potent compound against HDAC6, was exposed to anticancer evaluation against two human leukemia cancer cells; CCRF-CEM and MOLT-4 using MTT assay²⁷ (**Table 2**).

4. DISCUSSION

4.1 Chemistry

IR spectra for compounds **2a-c** showed sharp absorption bands ranging from 1780 to 1616 cm⁻¹ correlated with the presence of C=O groups. Whereas, thiohydantoin derivatives **2d-f** also clearly depicted C=S stretching at 1199, 1149 and 1180 cm⁻¹ respectively.

Moreover, the ¹HNMR analysis for **2a** revealed two doublets at δ 3.97 and δ 4.07 ppm ascertaining the presence of CH₂ protons in addition to a multiplet in the range of δ 4.72- 4.76 ppm indicating CH proton. ¹HNMR spectra for **2b,c** and **2d-f** showed two multiplets in the range of δ 3.61-4.01 and δ 4.56-4.82 ppm for CH₂ and CH protons, respectively.

Furthermore, ^{13}C NMR spectra of **2a-c** and **2d-f** exhibited two signals around δ 48.25 - δ 60.27 and δ 60.27 - δ 69.60 ppm, corresponding to the two aliphatic carbons CH_2 and CH , respectively.

4.2 HDAC6 assay:

4.2.1 Structure-activity relationship:

Like all reported HDAC inhibitors, the designed compounds have general characteristic structural features necessary for inhibiting HDAC6 enzymes. Herein, these pharmacophoric features represented by benzimidazole capping moiety bearing different substitution on 2-position, CH_2 linker and ZBG that contains either O or S atom for binding with Zn^{+2} ion in the enzyme active site.

Regarding substituents on 2-position of benzimidazole ring, the addition of methyl group in compound **2b** (IC_{50} = 166.09 ± 9.7 nM) resulted in an apparent increase in HDAC6 inhibition, comparing to unsubstituted analogue **2a** (IC_{50} = 279.59 ± 16 nM). Moreover, a 3-fold increase in HDAC6 inhibition was observed for phenyl benzimidazole derivative **2c** (IC_{50} = 97.35 ± 5.7 nM). Similarly, for thiohydantoin compounds **2d-f**, phenyl benzimidazole **2f** was much more potent than its unsubstituted (**2d**) and methyl (**2e**) analogues.

It is worth mentioning that the type of atom ($\text{X} = \text{O}, \text{S}$) on ZBG has an effective role in HDAC6 inhibitory activity of title compounds. It was noted that the hydantoin derivatives **2a,c** ($\text{X} = \text{O}$) were more potent than their corresponding thiohydantoin analogues **2d-f** ($\text{X} = \text{S}$).

4.3 In vitro anticancer activity:

Compound **2c** exhibited potent cytotoxic activity against the two tested cell lines showing one-digit micromolar IC_{50} s. Noticeably, compound **2c** (IC_{50} = 3.66 ± 0.22 μM) was 2-fold more active than SAHA reference drug (IC_{50} = 6.8 ± 0.41 μM) against MOLT-4 cell line. In addition, compound **2c** revealed half potency of SAHA against CCRF-CEM cells with IC_{50} s of 9.67 ± 0.58 and 3.83 ± 0.23 μM , respectively. It was also noted that MOLT-4 cell line was 2-fold more sensitive to compound **2c** than CCRF-CEM cell line.

4.4 Docking study:

In light of the well-established knowledge of the pharmacophoric parameters (cap, linker and ZBG) that are essential for HDAC6 inhibitory activity, a docking simulation was performed to clarify the interaction of the most promising compound (**2c**) with HDAC6. The molecular docking study was carried out using MOE 2014.0901 in HDAC6 isoenzyme that is co-crystallized with SAHA (PDB code: 5EEI)³⁰. As

speculated, compound **2c** was approached to the narrow channel of the binding site and chelated with Zn^{+2} ion at the bottom of catalytic site of HDAC6 (Fig 4a-e). In addition, compound **2c** formed three hydrogen bonds with the side chain residue His 574 and the backbone residue Gly 582 (Table 3). Furthermore, the 2-phenyl benzimidazole capping group formed additional hydrophobic contacts with important residues within the active pocket of HDAC6, including Leu 712, Ser 531, Phe 583, Phe 642, Phe 643, His 614, His 463 and Pro 464 that could contribute to significant improvement in compound activity. Accordingly, the removal or replacement of 2-phenyl benzimidazole with 2-methyl benzimidazole induces a great drop in HDAC6 inhibitory activity. This illustrates the important role of phenyl group in benzimidazole capping moiety. Moreover, CH_2 moiety of the linker was able to form an arene-H interaction with Phe 643. The obtained binding mode is observably matched with the pervious SAR study and could be considered as a valid explanation for the potent HDAC6 inhibition provoked by compound **2c**.

5. CONCLUSIONS

New 2-substituted benzimidazole (thio)hydantoin Mannich bases were synthesized as HDAC6 inhibitors. All derivatives inhibited HDAC6 at nanomolar level. Compound **2c** was the most potent and displayed HDAC6 inhibition equal to SAHA. In addition, **2c** was 2-fold more active than SAHA against MOLT-4 leukemia cells.

Funding statement: This work is not funded.

Conflict of interest: The authors declare no conflict of interest.

Authors consent of publication: This paper is published under the full approval of authors.

Author Contribution: All authors had full access to all the information and took responsibility for data integrity and data analysis accuracy. Authors H.S and H.G designed the study. Author R.M performed the experimental work. Authors H.G and R.M wrote the manuscript. Author H.S supervised the work and revised the whole manuscript. The final manuscript was read and accepted by all the contributors.

List of Abbreviations: SAHA (suberoylanilide hydroxamic acid), FDA (Food and Drug Administration); HDAC6 (histone deacetylase 6), ZBG (zinc binding group).

REFERENCES

- Kadioglu, O., Cao, J., Kosyakova, N., Mrasek, K., Liehr, T., & Efferth, T. (2016). Genomic and transcriptomic profiling of resistant CEM/ADR-5000 and sensitive CCRF-CEM leukaemia cells for unravelling the full complexity of multi-factorial multidrug resistance. *Scientific reports*, *6*, 36754. <https://doi.org/10.1038/srep36754>.
- Feinberg, A. P., & Tycko, B. (2004). The history of cancer epigenetics. *Nature reviews. Cancer*, *4*(2), 143–153. <https://doi.org/10.1038/nrc1279>.
- Heerboth, S., Lapinska, K., Snyder, N., Leary, M., Rollinson, S., & Sarkar, S. (2014). Use of epigenetic drugs in disease: an overview. *Genetics & epigenetics*, *6*, 9–19. <https://doi.org/10.4137/GEG.S12270>.
- Ell, B., & Kang, Y. (2013). Transcriptional control of cancer metastasis. *Trends in cell biology*, *23*(12), 603–611. <https://doi.org/10.1016/j.tcb.2013.06.001>.
- Seto, E., & Yoshida, M. (2014). Erasers of histone acetylation: the histone deacetylase enzymes. *Cold Spring Harbor perspectives in biology*, *6*(4), a018713. <https://doi.org/10.1101/cshperspect.a018713>.
- Li, T., Zhang, C., Hassan, S., Liu, X., Song, F., Chen, K., Zhang, W., & Yang, J. (2018). Histone deacetylase 6 in cancer. *Journal of hematology & oncology*, *11*(1), 111. <https://doi.org/10.1186/s13045-018-0654-9>.
- Mottamal, M., Zheng, S., Huang, T. L., & Wang, G. (2015). Histone deacetylase inhibitors in clinical studies as templates for new anticancer agents. *Molecules (Basel, Switzerland)*, *20*(3), 3898–3941. <https://doi.org/10.3390/molecules20033898>.
- Bi, G., & Jiang, G. (2006). The molecular mechanism of HDAC inhibitors in anticancer effects. *Cellular & molecular immunology*, *3*(4), 285–290.
- Kelly, W. K., & Marks, P. A. (2005). Drug insight: Histone deacetylase inhibitors--development of the new targeted anticancer agent suberoylanilide hydroxamic acid. *Nature clinical practice. Oncology*, *2*(3), 150–157. <https://doi.org/10.1038/ncponc0106>.
- Tambunan, U. S., Bramantya, N., & Parikesit, A. A. (2011). In silico modification of suberoylanilide hydroxamic acid (SAHA) as potential inhibitor for class II histone deacetylase (HDAC). *BMC bioinformatics*, *12 Suppl 13*(Suppl 13), S23. <https://doi.org/10.1186/1471-2105-12-S13-S23>.
- Butler, K. V., Kalin, J., Brochier, C., Vistoli, G., Langley, B., & Kozikowski, A. P. (2010). Rational design and simple chemistry yield a superior, neuroprotective HDAC6 inhibitor, tubastatin A. *Journal of the American Chemical Society*, *132*(31), 10842–10846. <https://doi.org/10.1021/ja102758v>.
- Ray, A., Das, D. S., Song, Y., Hideshima, T., Tai, Y. T., Chauhan, D., & Anderson, K. C. (2018). Combination of a novel HDAC6 inhibitor ACY-241 and anti-PD-L1 antibody enhances anti-tumor immunity and cytotoxicity in multiple myeloma. *Leukemia*, *32*(3), 843–846. <https://doi.org/10.1038/leu.2017.322>.
- Alpan, A. S., Zencir, S., Zupkó, I., Coban, G., Réthy, B., Gunes, H. S., & Topcu, Z. (2009). Biological activity of bis-benzimidazole derivatives on DNA topoisomerase I and HeLa, MCF7 and A431 cells. *Journal of enzyme inhibition and medicinal chemistry*, *24*(3), 844–849. <https://doi.org/10.1080/14756360802420831>.
- Zhang, J., Yao, D., Jiang, Y., Huang, J., Yang, S., & Wang, J. (2017). Synthesis and biological evaluation of benzimidazole derivatives as the G9a Histone Methyltransferase inhibitors that induce autophagy and apoptosis of breast cancer cells. *Bioorganic chemistry*, *72*, 168–181. <https://doi.org/10.1016/j.bioorg.2017.04.005>.
- Shrivastava, N., Naim, M. J., Alam, M. J., Nawaz, F., Ahmed, S., & Alam, O. (2017). Benzimidazole Scaffold as Anticancer Agent: Synthetic Approaches and Structure-Activity Relationship. *Archiv der Pharmazie*, *350*(6), 10.1002/ardp.201700040. <https://doi.org/10.1002/ardp.201700040>.
- Wang, T., Sepulveda, M., Gonzales, P., & Gately, S. (2013). Identification of novel HDAC inhibitors through cell based screening and their evaluation as potential anticancer agents. *Bioorganic & medicinal chemistry letters*, *23*(17), 4790–4793. <https://doi.org/10.1016/j.bmcl.2013.07.001>.

17. Cho, S., Kim, S. H., & Shin, D. (2019). Recent applications of hydantoin and thiohydantoin in medicinal chemistry. *European journal of medicinal chemistry*, 164, 517–545. <https://doi.org/10.1016/j.ejmech.2018.12.066>.
18. De Savi, C., Waterson, D., Pape, A., Lamont, S., Hadley, E., Mills, M., Page, K. M., Bowyer, J., & Maciewicz, R. A. (2013). Hydantoin based inhibitors of MMP13--discovery of AZD6605. *Bioorganic & medicinal chemistry letters*, 23(16), 4705–4712. <https://doi.org/10.1016/j.bmcl.2013.05.089>.
19. Kim, H. R., Lee, H. J., Choi, Y. J., Park, Y. J., Woo, Y., Kim, S. J., Moon, H. R. (2014). Benzylidene-linked thiohydantoin derivatives as inhibitors of tyrosinase and melanogenesis: importance of the β -phenyl- α,β -unsaturated carbonyl functionality. *Med. Chem. Commun.*, 5(9), 1410–1417. <https://doi.org/10.1039/c4md00171k>.
20. Preston, P. N. (1974). Synthesis, reactions, and spectroscopic properties of benzimidazoles. *Chemical Reviews*, 74(3), 279–314. <https://doi.org/10.1021/cr60289a001>.
21. Sharghi, H., Asemani, O., & Khalifeh, R. (2008). New One-Pot Procedure for the Synthesis of 2-Substituted Benzimidazoles. *Synthetic Communications*, 38(7), 1128–1136. <https://doi.org/10.1080/00397910701863657>.
22. Roman G. (2015). Mannich bases in medicinal chemistry and drug design. *European journal of medicinal chemistry*, 89, 743–816. <https://doi.org/10.1016/j.ejmech.2014.10.076>.
23. Tramontini, M., & Angiolini, L. (1990). Further advances in the chemistry of mannich bases. *Tetrahedron*, 46(6), 1791–1837 [https://doi.org/10.1016/s0040-4020\(01\)89752-0](https://doi.org/10.1016/s0040-4020(01)89752-0).
24. Popiołek, Ł., Rzymowska, J., Kosikowska, U., Hordyjewska, A., Wujec, M., & Malm, A. (2014). Synthesis, antiproliferative and antimicrobial activity of new Mannich bases bearing 1,2,4-triazole moiety. *Journal of enzyme inhibition and medicinal chemistry*, 29(6), 786–795. <https://doi.org/10.3109/14756366.2013.855926>.
25. Ozgun, D. O., Yamali, C., Gul, H. I., Taslimi, P., Gulcin, I., Yanik, T., & Supuran, C. T. (2016). Inhibitory effects of isatin Mannich bases on carbonic anhydrases, acetylcholinesterase, and butyrylcholinesterase. *Journal of enzyme inhibition and medicinal chemistry*, 31(6), 1498–1501. <https://doi.org/10.3109/14756366.2016.1149479>.
26. Rodríguez-Fonseca, R. A., Sixto-López, Y., Fragoso-Vázquez, M. J., Flores-Mejía, R., Cabrera-Pérez, L. C., Vázquez-Moctezuma, I., Rosales-Hernández, M. C., Bello, M., Martínez-Archundia, M., Trujillo-Ferrara, J. G., Becerra-Martínez, E., & Correa-Basurto, J. (2017). Design, Synthesis and Biological Evaluation of a Phenyl Butyric Acid Derivative, N-(4-chlorophenyl)-4-phenylbutanamide: A HDAC6 Inhibitor with Anti-proliferative Activity on Cervix Cancer and Leukemia Cells. *Anti-cancer agents in medicinal chemistry*, 17(10), 1441–1454. <https://doi.org/10.2174/1871520617666170103092851>.
27. Mosmann T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of immunological methods*, 65(1-2), 55–63. [https://doi.org/10.1016/0022-1759\(83\)90303-4](https://doi.org/10.1016/0022-1759(83)90303-4).
28. Mahmoud HK, Farghaly TA, Abdulwahab HG, Al-Qurashi NT, Shaaban MR. Novel 2-indolinone thiazole hybrids as sunitinib analogues: Design, synthesis, and potent VEGFR-2 inhibition with potential anti-renal cancer activity. *Eur J Med Chem*. 2020 Dec 15; 208:112752. <https://doi.org/10.1016/j.ejmech.2020.112752>.
29. Sami A. Al-Hussain, Thoraya A. Farghaly, Magdi E.A. Zaki, Hanan G. Abdulwahab, Nadia T. Al-Qurashi, Zeinab A. Muhammad Discovery of novel indolyl-1,2,4-triazole hybrids as potent vascular endothelial growth factor receptor-2 (VEGFR-2) inhibitors with potential anti-renal cancer activity *Bioorganic Chemistry* 105 (2020)104330. <https://doi.org/10.1016/j.bioorg.2020.104330>.
30. Hai, Y., & Christianson, D. W. (2016). Histone deacetylase 6 structure and molecular basis of catalysis and inhibition. *Nature chemical biology*, 12(9), 741–747. <https://doi.org/10.1038/nchembio.2134>.