

Synthesis, biological assessment, and docking study of new pyrazolo[1,5-*a*]pyrimidine derivatives with potential anticancer activity

1. Experimental

1.1. Chemistry

All of the chemicals and solvents were commercially available, supplied from Aldrich (Germany), Merk (Germany), Loba Chemie (India) and used without any further purification. Melting points were measured in open capillary tubes using Griffin apparatus and were uncorrected apparatus. Elemental Microanalysis was carried out at the Regional Center for Mycology and Biotechnology, Al-Azhar University. The monitoring of the progress of all reactions and homogeneity of the synthesized compounds was carried out by thin layer chromatography (TLC) using Silica gel-precoated aluminum sheets (type 60, F 254 0.2mm thickness, Merck, Darmstadt, Germany). Visualization of compound spots on TLC and follow up were made using UV lamp at λ 254 nanometer. Infrared IR spectra were recorded using potassium bromide disc technique on a Shimadzu 435 IR spectrophotometer at October 6 University. Mass Spectra were recorded using Shimadzu Gas Chromatograph Mass spectrometer-Qp 2010 plus (Japan) at the Regional Center for Mycology and Biotechnology, Al-Azhar University. ^1H NMR and ^{13}C NMR spectra were recorded on a Varian-Mercury 400 (^1H , 400 MHz; ^{13}C , 400 MHz) spectrometer at the DNA Research Center, Faculty of Science, Zagazig University and Bruker high-performance digital FT-NMR spectrometer AVANCE III 400 MHz (Bruker Corporation, Germany) at the Microanalytical Unit, Faculty of Pharmacy, Cairo University using DMSO-*d*₆ as a solvent. δ Parts per million (ppm) are the chemical shifts quotations downfielding from tetramethylsilane (TMS) as an internal standard. All chemical shifts were reported in ppm, the coupling constants (*J*) were expressed in hertz (Hz) and signals (multiplicities) were described as singlet (s), doublet (d), triplet (t), quartet (q) as well as multiplet (m).

1.2. Biology

MCF-7 (breast), HepG2 (liver) and A549 (lung) cancer cell lines were grown as monolayer and routinely maintained in RPMI-1640 medium supplemented with 5% heat inactivated FBS, 2 mM glutamine and antibiotics (penicillin 100 U/ml, streptomycin 100 mg/ml) at 37° C in a humidified atmosphere containing 5% CO₂. Exponentially growing cells were obtained by plating 1.5 x10⁵ cells/ml, followed by 24 h of

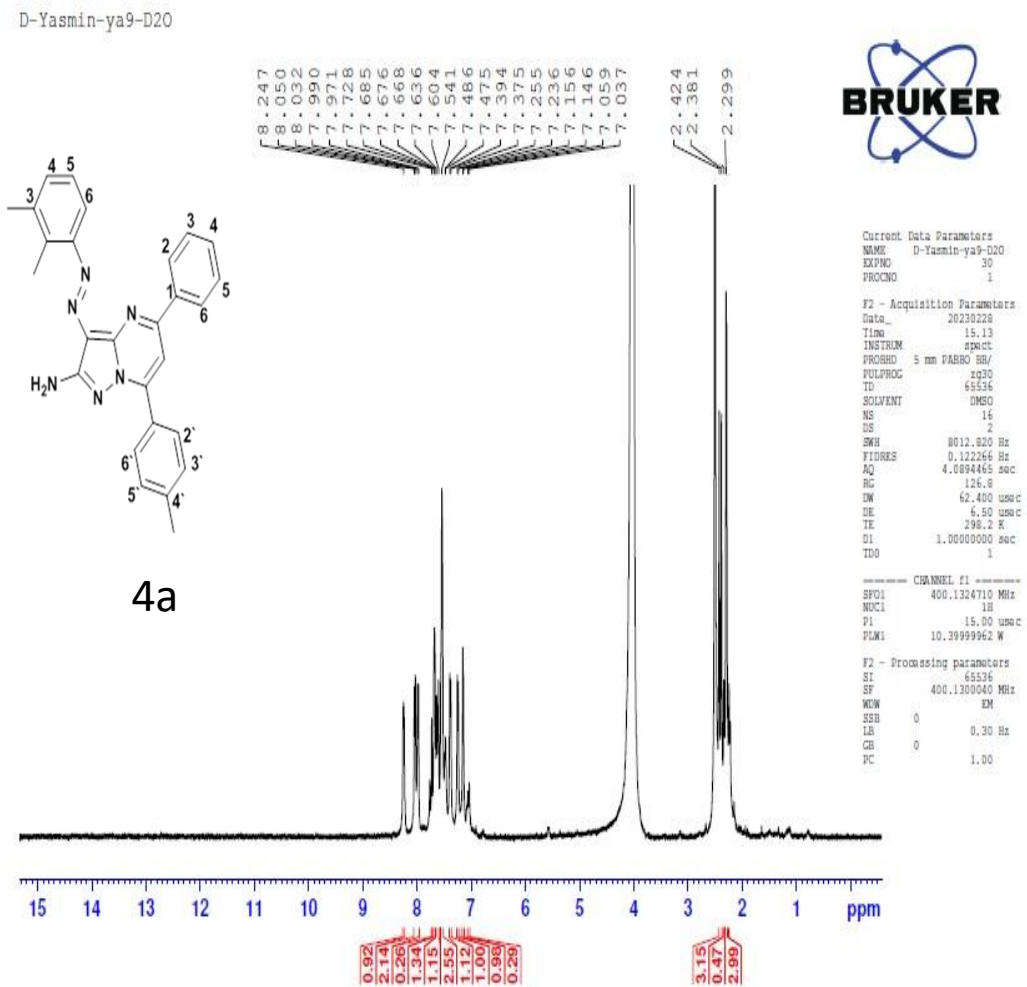
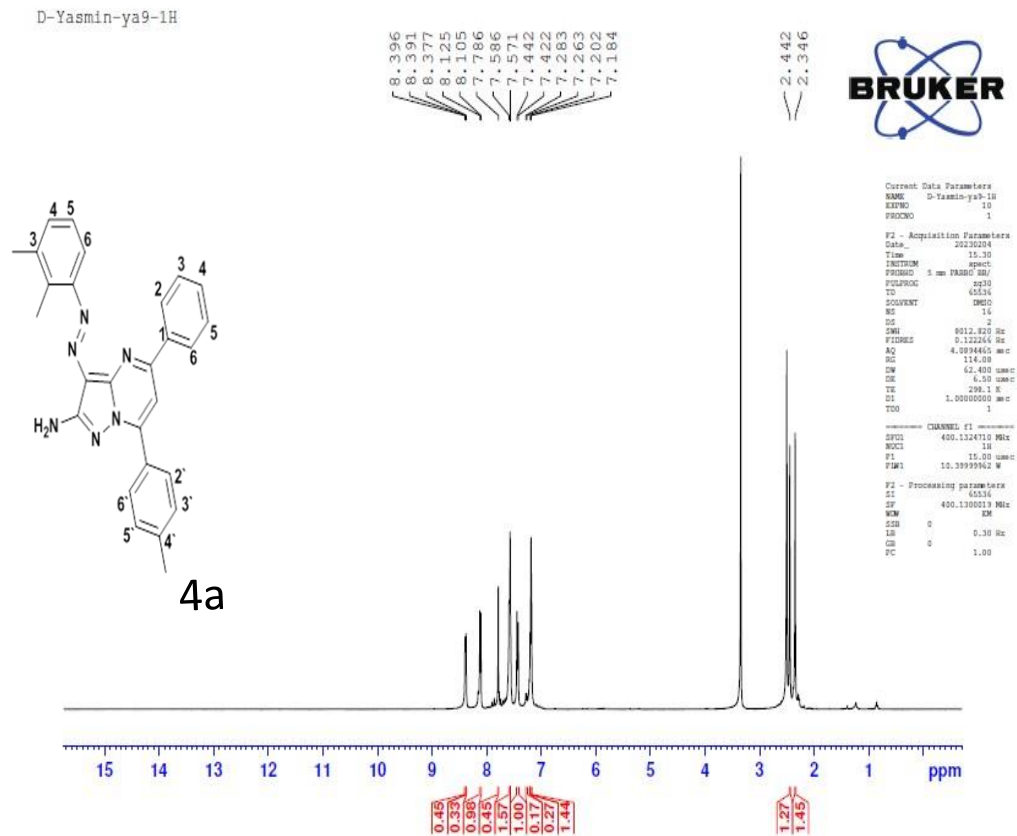
incubation. The effect of the vehicle solvent DMSO on the growth of these cell lines was evaluated in all the experiments by exposing untreated control cells to the maximum concentration (0.5%) of DMSO used in each assay. The activity of the compounds was evaluated using sulforhodamine B dye where cells were treated with five different serial dilutions (up to 150 mM) of the compounds for 48 h. Doxorubicin was used as a positive control and tested in the same manner. The cytotoxicity of the compounds were then determined using the SRB method as previously described by Skehan *et al* ¹. The IC₅₀ values were calculated according to the equation for Boltzman sigmoidal concentration response curve using the nonlinear regression fitting models (Graph Pad, Prism Version 5).

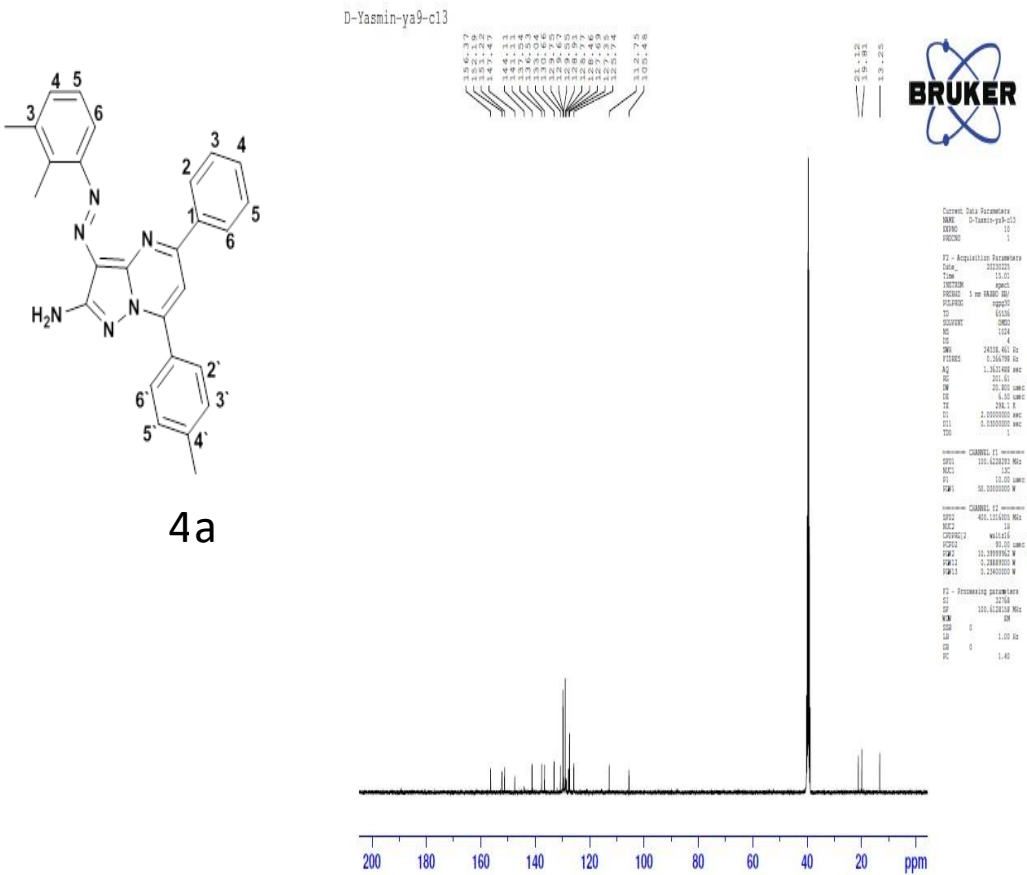
1.3. Docking

Molecular Operating Environment (MOE) software model 2019.01 version was used for performing the molecular docking studies. The X-ray crystal structure of VEGFR-2 in a complex with sorafenib was downloaded from <https://www.rcsb.org/> (PDB ID: 4ASD). The protein-ligand complex bought from the protein data bank was organized for docking: (1) chain B of the protein along with co-crystallized water molecules were deleted; (2) the enzyme was 3D protonated and (3) sorafenib was redocked into the active site for validation of the docking procedure. Scoring was performed using the Alpha triangle placement technique and London dG. Thirty of the most stable docking poses for ligand were retained with the best scoring conformation was done. The validation results confirmed a near ideal alignment with the original ligand and displayed the same binding interactions as obtained from the X-ray crystallography pdb document. Moreover **4d** compound was drawn on the MOE. A Hamiltonian-Force Field-MMFF94x was used to reduce structure energy and it was docked into the active site of enzyme. The most stable ten conformers for compound have been retained.

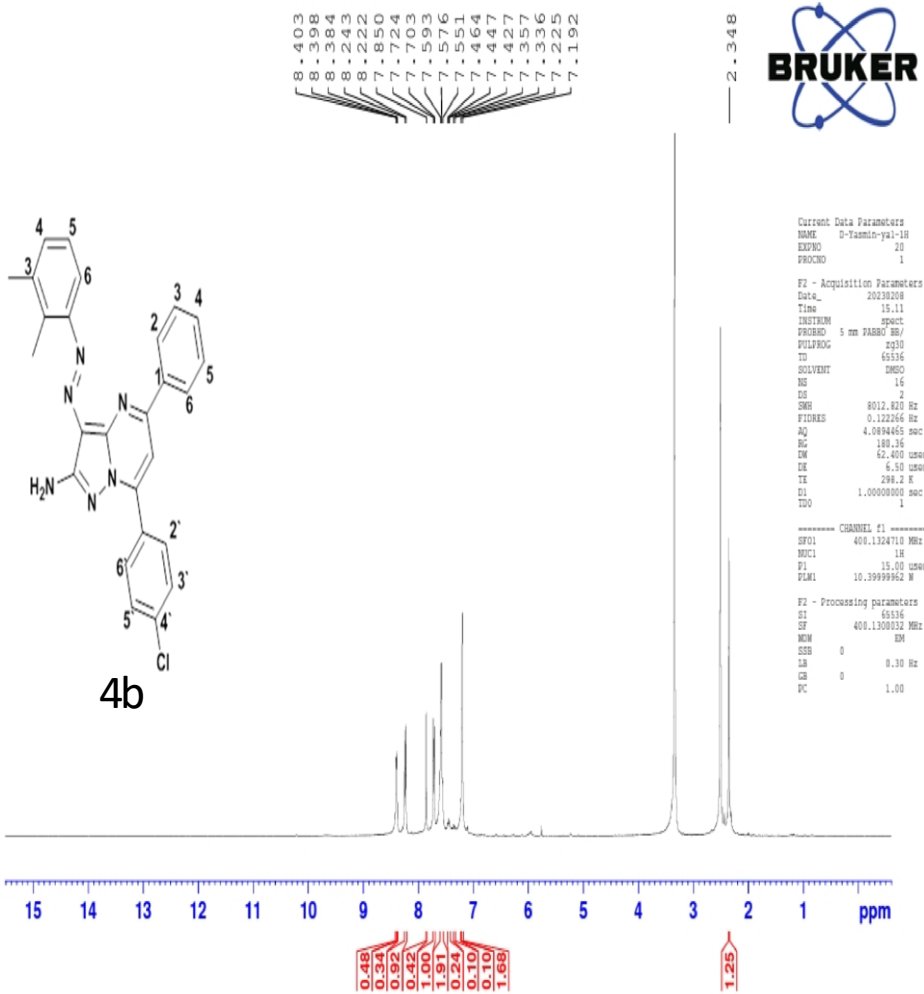
REFERENCES

1. Skehan P, Storeng R, Scudiero D, Monks A, McMahon J, Vistica D, et al. New colorimetric cytotoxicity assay for anticancer-drug screening. *J. Natl. Cancer Inst.* 1990; 82: 1107-1112.

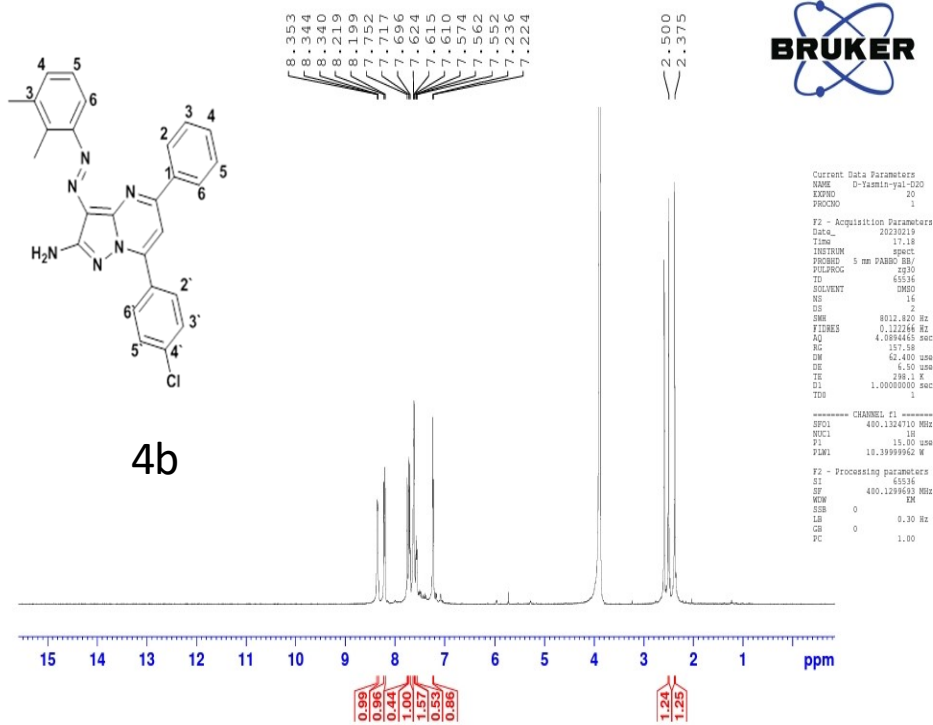




D-Yasmin-yal-1H

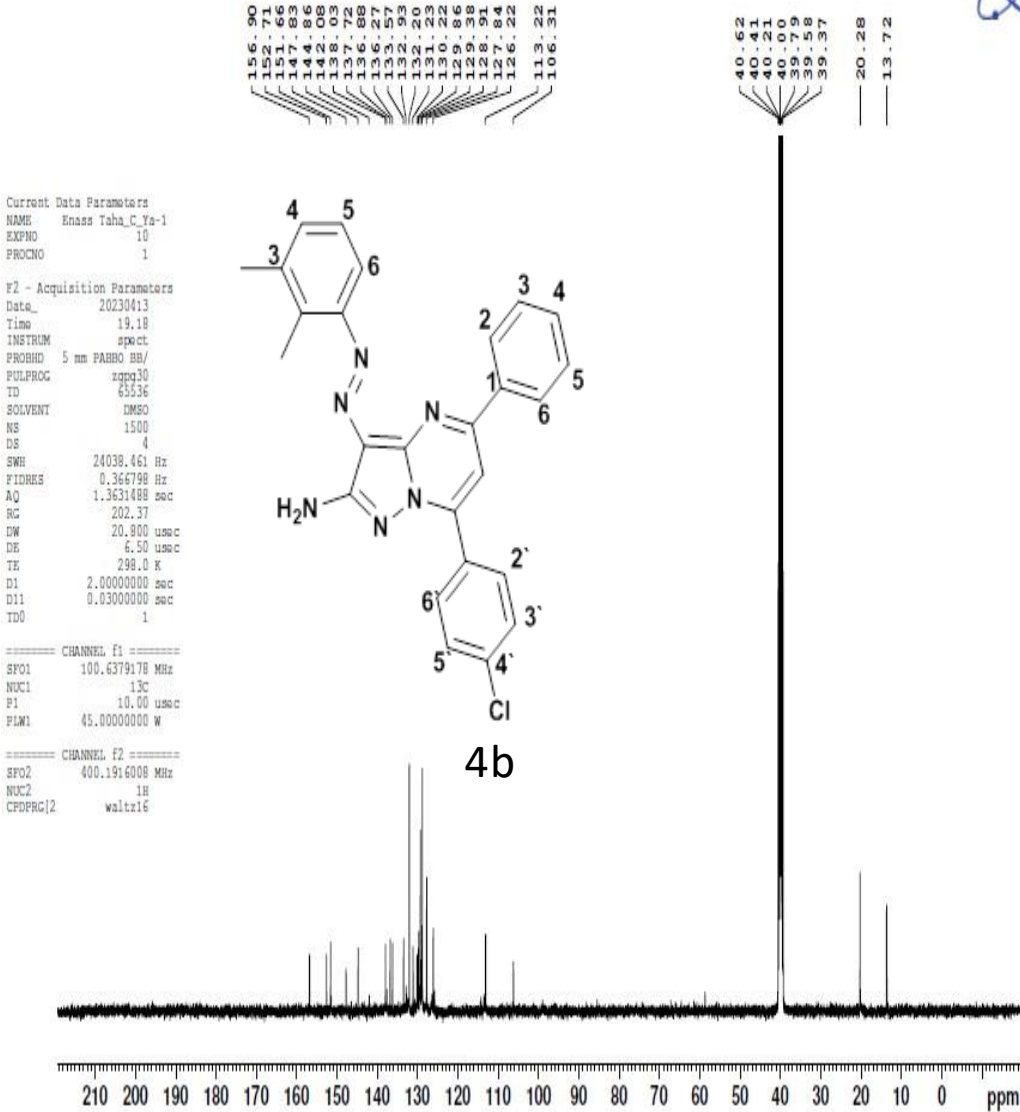


D-Yasmin-yal-D2O

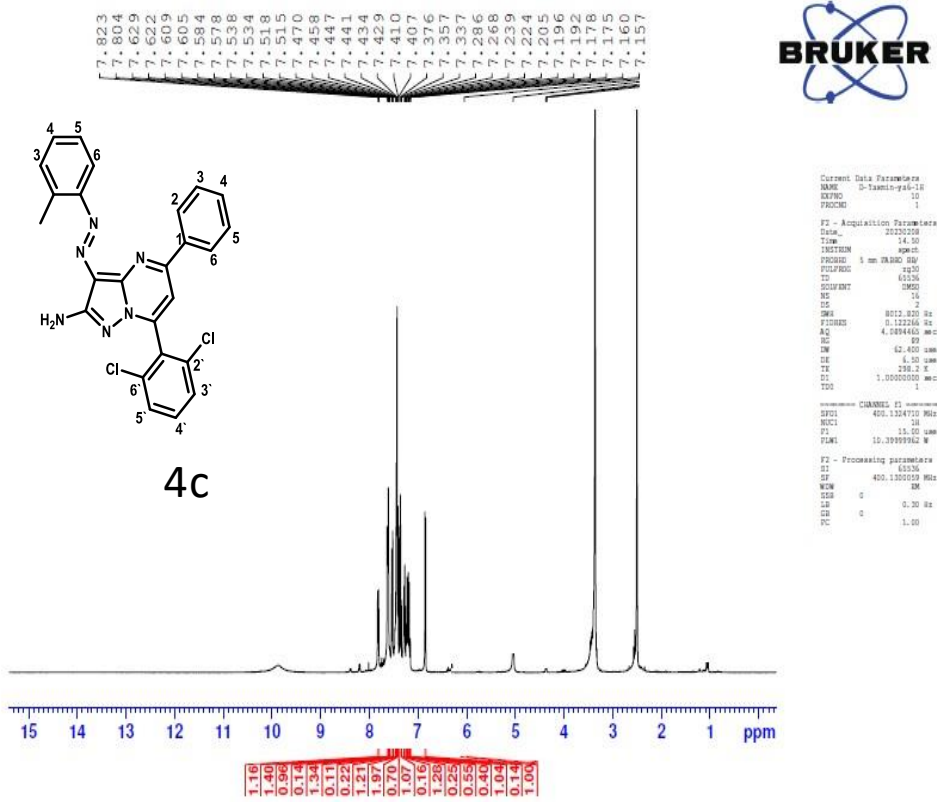


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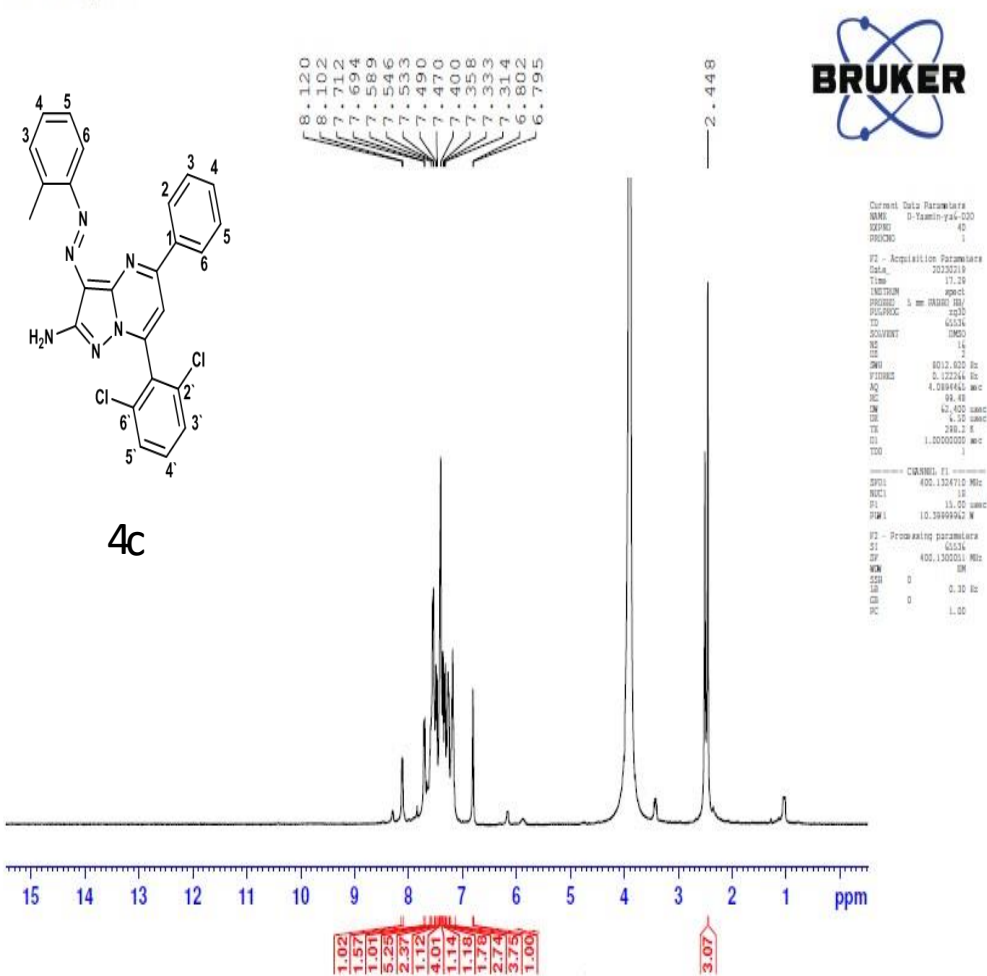
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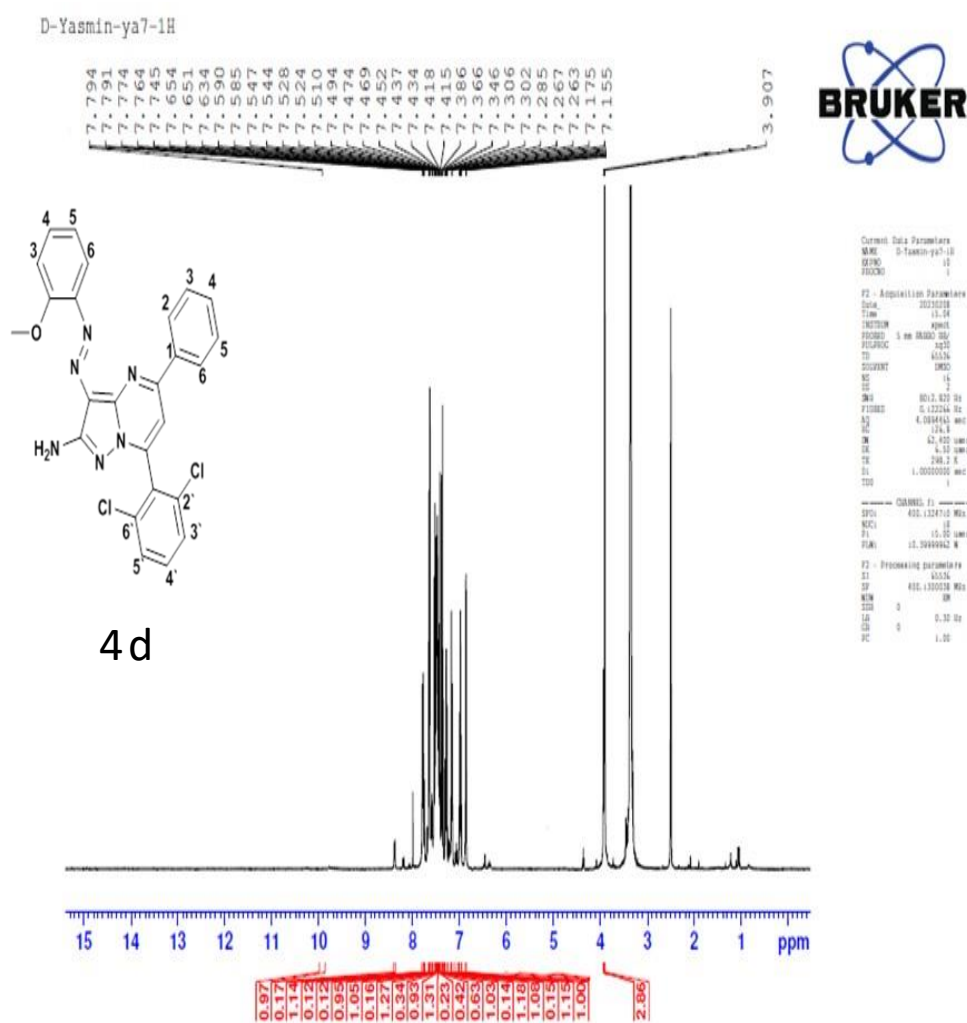
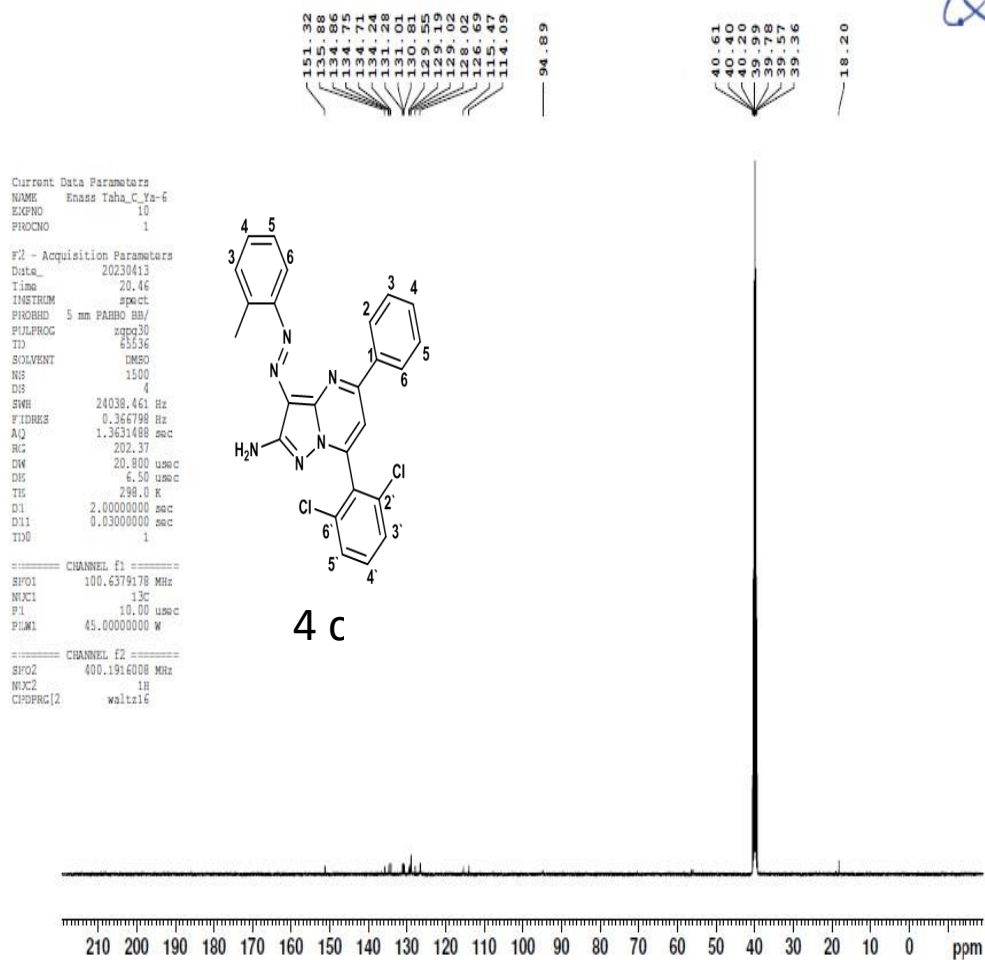


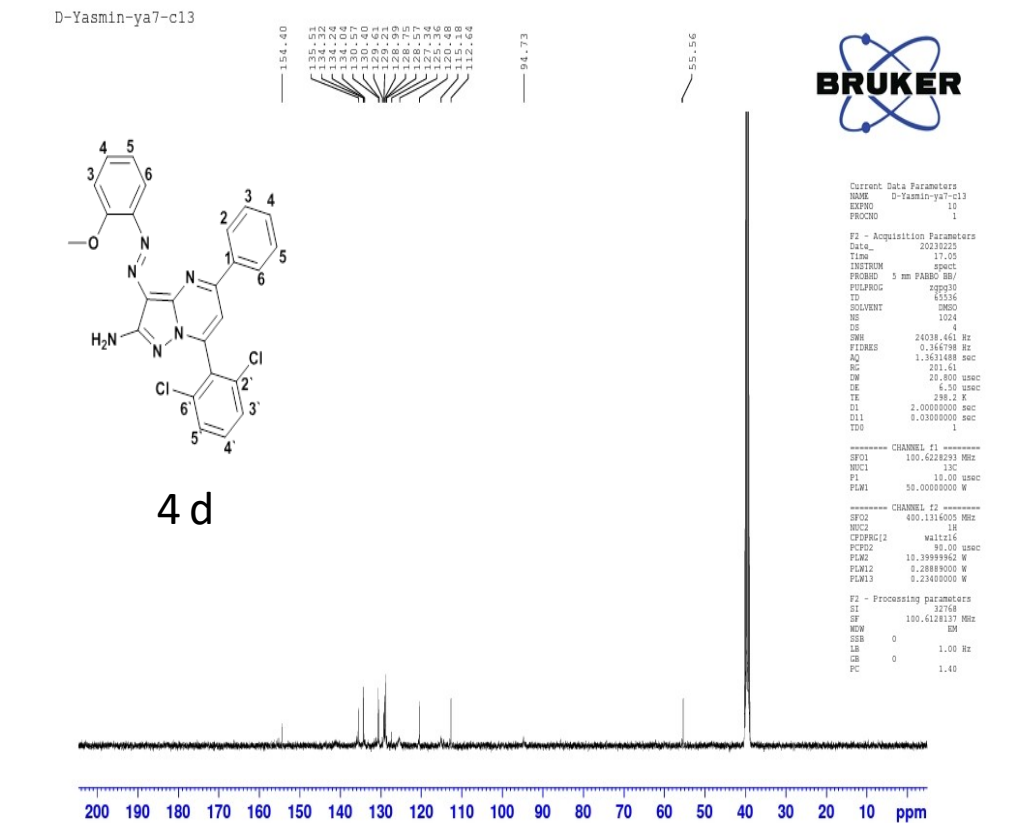
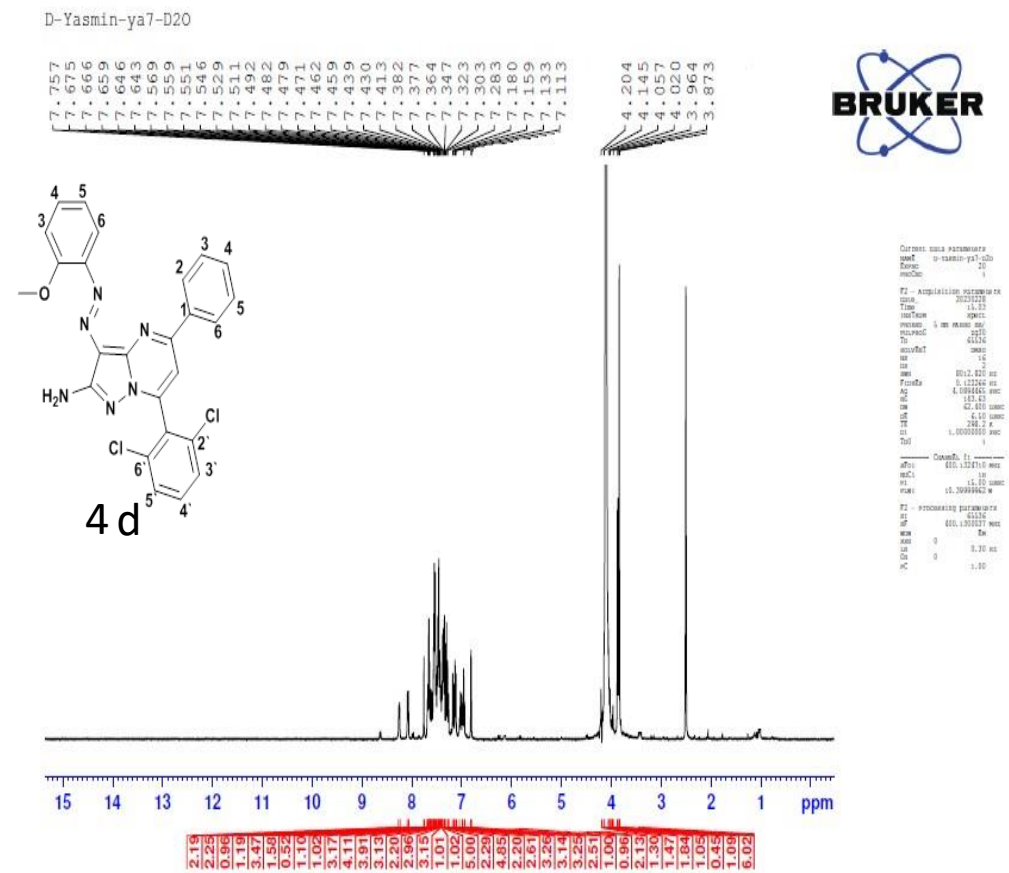
D-Yasmin-ya6-1H



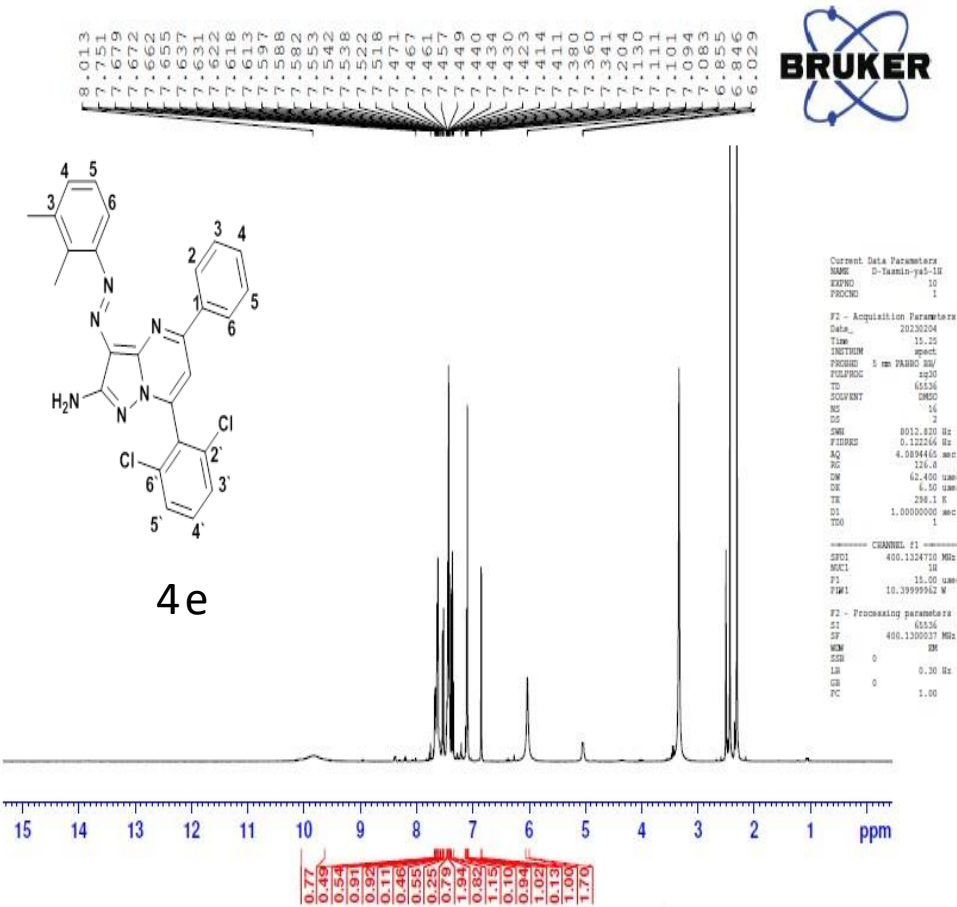
D-Yasmin-ya6-D2O







D-Yasmin-ya5-1H



D-Yasmin-ya5-D2O

